UNITED STATES ENVIRONMENTAL PROTECTION AGENCY WASHINGTON D.C. 20460

August 13, 1998

OFFICE OF THE ADMINISTRATOR SCIENCE ADVISORY BOARD

Note to the Reader:

The attached draft report is a draft report of the Science Advisory Board (SAB). The draft is still undergoing final internal SAB review, however, in its present form, it represents the consensus position of the panel involved in the review. Once approved as final, the report will be transmitted to the EPA Administrator and will become available to the interested public as a final report.

This draft has been released for general information to members of the interested public and to EPA staff. This is consistent with the SAB policy of releasing draft materials only when the Committee involved is comfortable that the document is sufficiently complete to provide useful information to the reader. The reader should remember that this is an unapproved working draft and that the document should not be used to represent official EPA or SAB views or advice. Draft documents at this stage of the process often undergo significant revisions before the final version is approved and published.

The SAB is not soliciting comments on the advice contained herein. However, as a courtesy to the EPA Program Office which is the subject of the SAB review, we have asked them to respond to the issues listed below. Consistent with SAB policy on this matter, the SAB is not obligated to address any responses which it receives.

- 1. Has the Committee adequately responded to the questions posed in the Charge?
- 2. Are any statements or responses made in the draft unclear?
- 3. Are there any technical errors?

For further information or to respond to the questions above, please contact:

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United States

Environmental

Protection Agency

AN SAB REPORT: REVIEW OF THE HEALTH RISK ASSESSMENTOF 1,3-BUTADIENE

REVIEW OF THE OFFICE OF RESEARCH AND DEVELOPMENT'S DRAFT HEALTH RISK ASSESSMENT OF 1,3-BUTADIENE PREPARED BY THE ENVIRONMENTAL HEALTH COMMITTE (EHC)

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EPA-SAB-EHC-98-

Honorable Carol M. Browner

Administrator

U.S. Environmental Protection Agency

401 M Street, S.W.

Washington, DC 20460

Subject: Review of the Health Risk Assessment of 1,3-Butadiene

Dear Ms. Browner:

At the request of the Office of Research and Development (ORD), the Environmental Health Committee (EHC) of the Environmental Protection Agency's Science Advisory Board (SAB) reviewed the Agency's Health Risk Assessment of 1,3-butadiene. The Committee met on April 30 and May 1, 1998 at the EPA's Waterside Mall Complex in Washington DC. The EHC approved the Committee's report on August 11, 1998 and the SAB's Executive Committee approved this report on ______.

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The Health Risk Assessment of 1,3-butadiene which was reviewed by the Committee was developed by the Office of Research and Development. ORD published its first risk assessment of 1,3-butadiene in 1985. The first document covered cancer and mutagenicity and was prepared in response to a request from the Office of Air Quality Planning and Standards to support the

classification of 1,3-butadiene as a Hazardous Air Pollutant. The recent draft 1,3-butadiene		
document was written in response to a request from the Agency's Office of Mobile Sources. The		
Agency plans to use the final document to support a future Air Toxics Rule. The recent draft		
1,3-butadiene document (the review document) focuses on mutagenicity, carcinogenicity, and		
reproductive/developmental effects. The 1,3-butadiene document which was reviewed by the		
Committee presents the Agency's first benchmark dose analysis for reproductive/developmental		
factors. The review document includes many new studies which have been published since 1985.		
The EPA concludes, in its draft health risk assessment of 1,3-butadiene, that this new information		
has changed the weight of evidence for cancer. In addition, there are exposure data available in ar		
occupational study which are used to derive the cancer slope factor. The review document is not		
intended to be a comprehensive health assessment. For example, it contains an overview of the		
ambient exposure and exposure of populations adjacent to emissions sources, without any actual		
exposure assessment as such.		
In addition to a general review of the document by the Committee, the Office of Research		
and Development specifically requested that the EHC provide comment on each of the following		
aspects of the document:		
a) Review the health risk assessment for technical quality, comprehensiveness and clarity		
(Address each chapter, but with specific reference to Charges b, c, and d).		
b) Does the science support the classification of "known" human carcinogen?		
c) Are the approaches taken to characterize plausible cancer risks reasonable given the		
science?		
d) Are the conclusions and quantitative estimations for reproductive/developmental		

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effects adec	uately s	supported?
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The EHC recognizes that preparing a health risk assessment of 1,3-butadiene was a difficult and complicated task given the large amount of information on 1,3-butadiene toxicity, epidemiology, and mechanisms available, with new and relevant information being made on a continual basis. Furthermore, there has been a significant body of literature published since the last carcinogen assessment for 1,3-butadiene which was conducted by the EPA in 1985.

The Agency selected a cutoff date for the inclusion of new information in the revised, draft document of January 31, 1997. However, a significant amount of new and important information has been developed since then and is pertinent to a health risk assessment of 1,3-butadiene. The Committee felt that the report should reflect the most current research data, including the recent evaluations by the International Agency for Research on Cancer, and if finalized soon, the evaluation of Health Canada (IARC, 1998)(Health Canada, 1997). Therefore, with respect to the first charge question on the technical quality, comprehensiveness and clarity of the document, the EHC found that the quality and comprehensiveness would be greatly improved by including research data published in the peer-reviewed literature since the cutoff date of January 31, 1997. The EHC notes that important research such as the Delzell et al. (1995) exposure reestimation and the pharmacokinetic modeling studies is ongoing. To improve the clarity of the document, the Committee recommended several editorial changes, such as the inclusion of summary tables in some of the chapters.

The majority of the Environmental Health Committee did not classify 1,3-butadiene as a known human carcinogen due to the lack of consistency between exposure and leukemia or lymphosarcoma data when the styrene-butadiene rubber (SBR) and monomer worker studies were considered in total. The majority opined that 1,3-butadiene should be classified as a probable human carcinogen whereas a minority felt that the science supported the classification

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of 1,3-butadiene as a known human carcinogen. The vast majority of the panel felt that the SBR process supported a classification as a known carcinogen. The majority opinion was based on several lines of evidence: a) There was only one study on workers in the monomer process that showed an excess of lymphosarcoma but this study has not been replicated. (One would like to see at least a second independent confirmatory study before affirming that there is "sufficient evidence of human carcinogenicity" regarding butadiene and leukemia. Instead a fairly large and reasonable second study shows no leukemia excess and two smaller ones found no evidence of leukemia risk); b) In the monomer study, there was no evidence of an exposure response relationship for lymphosarcoma nor was there evidence that those workers with longer-term exposure had a higher risk of lymphosarcoma; and c) A large study of the workers in the styrene-butadiene rubber (SBR) industry showed an excess of leukemia. However, since these workers were exposed to several different chemicals, the cancer excess could not be attributed solely to 1,3-butadiene. Furthermore, the findings from the reevaluation of the exposure estimates in the Delzell et al. (1995) study may impact the risk assessment. The majority of the Committee felt that the finding of 'known human carcinogen' should solely be based on observational studies in humans, without regard to mechanistic or other information. Others on the Committee felt that butadiene should be identified as a 'known human carcinogen' because the cumulative evidence, from epidemiology, animal cancer bioassays, and mechanistic studies should be used as the basis for the judgment.

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The Committee found the approaches taken to characterize plausible cancer risks to be reasonable but points out specific data that may have been misinterpreted by the Agency. In particular, the discussion of metabolism and toxicokinetics failed to address critical differences in metabolites of butadiene in different species accounting for differences in tumor susceptibility in different species. Inclusion of a discussion of state-of-the-art models for butadiene kinetics would significantly strengthen the scientific quality of the document. Included in this discussion should be the strengths and weaknesses of the available models for risk prediction. This fuller

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1	discussion sho	ould replace the simple statements made about the inadequacy of the available
2	models.	
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4	The Co	ommittee commends the Agency for looking at new approaches, such as the
5	benchmark do	se procedure, to improve quantitative assessment of non cancer endpoints.
6	However, the	Committee has submitted suggestions on how to further improve these approaches
7	and how to ma	ake these new approaches more clear, accurate and concise, including the following
8	recommendati	ons:
9	a)	There was an apparent mathematical error in the calculation of the benchmark
10		concentration for reproductive and developmental effects which must be
11		addressed. All calculations should be easy to follow, and of course, carefully
12		proofed;
13	b)	There are new dominant lethal studies, which are not included in the risk
14		assessment, that failed to replicate earlier findings;
15	c)	A variety of viewpoints were expressed within the Committee over the extent to
16		which toxicokinetic analyses should be incorporated into the assessment. At a
17		minimum the different toxicokinetic hypotheses and the hypothesized role of the
18		various metabolites should be discussed, at least qualitatively, with an indication of
19		the degree to which the assessment would be impacted if some of the hypotheses
20		were later proven true.
21	d)	Inadequate justification is given for the application of the additional safety factor
22		for the benchmark dose. Some of the Committee members could not understand
23		the rationale for its inclusion, and
24	e)	The rationale for the selection of the toxic non-cancer endpoint that is utilized in
25		the derivation of the RfC is very important and should be more explicitly

explained.

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1	The Committee appreciates the opportunity to review the draft Health Risk Assessment of
2	1,3-butadiene and looks forward to receiving a written response from the Director of the Office of
3	Research and Development, National Center for Environmental Assessment.
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6	Sincerely,
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8	Dr. Joan M. Daisey, Chair
9	Executive Committee
10	
11	Dr. Emil A. Pfitzer, Chair
12	Environmental Health Committee
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14	Mark J. Utell, Co-Chair,
15	Environmental Health Committee
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This report has been written as part of the activities of the Science Advisory Board, a public advisory group providing extramural scientific information and advice to the Administrator and other officials of the Environmental Protection Agency. The Board is structured to provide balanced, expert assessment of scientific matters related to problems facing the Agency. This report has not been reviewed for approval by the Agency and, hence, the contents of this report do not necessarily represent the views and policies of the Environmental Protection Agency, nor of other agencies in the Executive Branch of the Federal government, nor does mention of trade names or commercial products constitute a recommendation for use.

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ABSTRACT

The Environmental Health Committee (EHC) reviewed the EPA's updated, draft health risk assessment of 1,3-butadiene. The Committee acknowledged that this is an extremely difficult task given the large amount of information on 1,3-butadiene toxicity, epidemiology, and mechanisms, with new information being introduced on a continual basis.

The Agency selected a cutoff date for the inclusion of new information in the revised, draft document of January 31, 1997. However, a significant amount of new and important information has been developed since then and is pertinent to a health risk assessment of 1,3-butadiene. The Committee felt that the report should reflect the most current research data, including the recent evaluations by the International Agency for Research on Cancer, and if finalized soon, the evaluation of Health Canada. The EHC notes that important research such as the Delzell et al. (1995) exposure reestimation and the pharmacokinetic modeling studies is ongoing. To improve the clarity of the document, the Committee recommended several editorial changes, such as the inclusion of summary tables in some of the chapters.

The majority of the Environmental Health Committee did not classify 1,3-butadiene as a known human carcinogen due to the lack of consistency between exposure and leukemia or lymphosarcoma data when the styrene-butadiene rubber (SBR) and monomer worker studies were considered in total. The majority opined that 1,3-butadiene should be classified as a probable human carcinogen where as a minority felt that the science supported the classification of 1,3-butadiene as a known human carcinogen. The vast majority of the panel felt that the SBR process supported classification as a known carcinogen. The majority opinion was based on several lines of evidence: a) There was only one study of workers in the monomer process that showed an excess of lymphosarcoma but this study has not been replicated; b) In the monomer study, there was no evidence of an exposure response relationship for lymphosarcoma nor was

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there evidence that those workers with longer-term exposure had a higher risk of lymphosarcoma; and c) A large study of the workers in the styrene-butadiene rubber (SBR) industry showed an excess of leukemia. However, since these workers were exposed to several different chemicals, the cancer excess could not be attributed solely to 1,3-butadiene.

The Committee found the approaches taken to characterize plausible cancer risks to be reasonable but points out specific data that may have been misinterpreted by the Agency. In particular, the discussion of metabolism and toxicokinetics failed to address critical differences in metabolites of butadiene in different species accounting for differences in tumor susceptibility. Inclusion of a discussion of state-of-the-art models for butadiene kinetics would significantly strengthen the scientific quality of the document. Included in this discussion should be the strengths and weaknesses of the available models for risk prediction. This fuller discussion should replace the simple statements made about the inadequacy of the available models.

The Committee supported the use of the benchmark dose procedure in developing Reference levels. The Committee submitted suggestions on how to further improve the approaches for quantitative assessment of non-cancer endpoints, such as the reproductive/developmental effects, and how to make the approach more clear, accurate and consistent. All calculations should be easy to follow, and of course, carefully proofed. Greater explanation is needed of the safety factors applied to the benchmark, and of the newly proposed models, especially those modeling time to impact. New dominant lethal studies, not included in the risk assessment should be evaluated, especially in light of apparent inconsistency with the earlier findings. The Agency should consider a quantitative assessment based on the heritable translocation endpoint. Also, the EHC recommends that the Agency explain, in more detail, the rationale for the selection of the toxic non-cancer endpoint that is utilized in the derivation of the RfC.

1	<u>Keywords</u> :	1,3-butadiene, EPA's proposed Cancer Risk Assessment Guidelines, known human
2		carcinogen, probable human carcinogen, lymphosarcoma, leukemia,
3		reproductive/developmental effects, pharmacokinetics, risk assessment
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	ENVIRONMENTAL HEALTH COMMITTEE
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1. EXECUTIVE SUMMARY

The EPA Office of Research and Development prepared the draft *Health Risk Assessment* of 1,3-Butadiene (USEPA, 1998a) in response to a request from the Office of Mobile Sources (OMS). The document was requested by OMS in order to support a future Air Toxics Rule. The review document is not intended to be a comprehensive health assessment and therefore, does not contain any actual exposure assessment. The document focuses on mutagenicity, carcinogenicity, and reproductive/developmental effects and presents the Agency's first benchmark dose analysis for reproductive/developmental effects. In its document, the Agency has found that the new studies which have been published since 1985 change the weight of evidence for cancer. Based on the weight of overall evidence from human, animal, and mutagenicity studies, the Agency concludes that 1,3-butadiene is a known human carcinogen.

On April 30 and May 1, 1998, the Environmental Health Committee met at the EPA's Waterside Mall complex in Washington, DC to review the Agency's draft *Health Risk Assessment of 1,3-Butadiene*. In addition to a general review of the document by the Committee, the Office of Research and Development specifically requested that the EHC provide comment on each of the following aspects of the document:

a) Review the health risk assessment for technical quality, comprehensiveness and clarity (Address each chapter, but with specific reference to Charges b, c, and d).

b) Does the science support the classification of "known" human carcinogen?

c) Are the approaches taken to characterize plausible cancer risks reasonable given the science?

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d) Are the conclusions and quantitative estimations for reproductive/developmental effects adequately supported?

The Committee acknowledges that this is an extremely difficult task given the large amount of information on 1,3-butadiene toxicity, epidemiology, and mechanism available with new information being made on a continual basis.

The Agency selected a cutoff date for the inclusion of new information in the revised, draft document of January 31, 1997. However, a significant amount of new and important information has been developed since then and is pertinent to a health risk assessment of 1,3-butadiene. The Committee felt that the report should reflect the most current research data, including the recent evaluations by the International Agency for Research on Cancer and on ongoing evaluation in Canada (Health Canada, 1997)(IARC, 1998). Therefore, with respect to the first charge question on the technical quality, comprehensiveness and clarity of the document, the EHC found that the quality and comprehensiveness would be greatly improved by including research data published in the peer-reviewed literature since the cutoff date of January 31, 1997. The EHC notes that important research such as the Delzell et al. (1995) exposure reestimation and the pharmacokinetic modeling studies is ongoing. To improve the clarity of the document, the Committee recommended several editorial changes, such as the inclusion of summary tables in some of the chapters.

The majority of the Environmental Health Committee did not classify 1,3-butadiene as a known human carcinogen due to the lack of consistency between exposure and leukemia or lymphosarcoma data when the styrene-butadiene rubber (SBR) and monomer worker studies were considered in total. The majority opined that 1,3-butadiene should be classified as a probable human carcinogen where as a minority felt that the science supported the classification of

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1,3-butadiene as a known human carcinogen. The vast majority of the panel felt that the SBR process supported a classification as a known carcinogen. The majority opinion was based on several lines of evidence: a) There was only one study on workers in the monomer process that showed an excess of lymphosarcoma but this study has not been replicated. (One would like to see at least a second independent confirmatory study before affirming that there is "sufficient evidence of human carcinogenicity" regarding butadiene and leukemia. Instead a fairly large and reasonable second study shows no leukemia excess and two smaller ones found no evidence of leukemia risk); b) In the monomer study, there was no evidence of an exposure response relationship for lymphosarcoma nor was there evidence that those workers with longer-term exposure had a higher risk of lymphosarcoma; and c) A large study of the workers in the styrene-butadiene rubber (SBR) industry showed an excess of leukemia. However, since these workers were exposed to several different chemicals, the cancer excess could not be attributed solely to 1,3-butadiene. The majority of the Committee felt that the finding of 'known human carcinogen' should solely be based on observational studies in humans, without regard to mechanistic or other information. Others on the Committee felt that butadiene should be identified as a 'known human carcinogen' because the cumulative evidence, from epidemiology, animal cancer bioassays, and mechanistic studies should be used as the basis for the judgment.

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The Committee commends the Agency for looking at new approaches, such as the		
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However, the	Committee has submitted suggestions on how to further improve these approaches	
and how to m	nake these new approaches more clear, accurate and concise, including the following	
recommendat	ions:	
a)	There was an apparent mathematical error in the calculation of the benchmark	
	concentration for reproductive and developmental effects which must be	
	addressed. All calculations should be easy to follow, and of course, carefully	
	proofed;	
b)	There are new dominant lethal studies, which are not included in the risk	
	assessment, that failed to replicate earlier findings;	
c)	A variety of viewpoints were expressed within the Committee over the extent to	
	which toxicokinetic analyses should be incorporated into the assessment. At a	
	minimum the different toxicokinetic hypotheses and the hypothesized role of the	
	various metabolites should be discussed, at least qualitatively, with an indication of	
	the degree to which the assessment would be impacted if some of the hypotheses	
	were later proven true.	
d)	Inadequate justification is given for the application of the additional safety factor	
	for the benchmark dose. Some of the Committee members could not understand	
	the rationale for its conclusion, and	
e)	The rationale for the selection of the toxic non-cancer endpoint that is utilized in	
	the derivation of the RfC is very important and should be more explicitly	
	explained.	

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2. INTRODUCTION

2.1 Background

The draft *Health Risk Assessment of 1,3-Butadiene* (USEPA, 1998a) was developed by the EPA Office of Research and Development at the request of the Office of Mobile Sources. This document was requested to support a future Air Toxics Rule. The document was not intended to be a comprehensive health assessment. Consequently, an actual exposure assessment is not included in the document. The draft *Health Risk Assessment of 1,3-Butadiene* (USEPA, 1998a) focuses on carcinogenicity, mutagenicity, and reproductive/developmental effects. The document presents the Agency's first benchmark dose analysis for reproductive/developmental factors.

The draft document that was reviewed by the Committee updates a previously, published document (USEPA, 1995). In the current draft document, the Agency has included many new studies which have been published since 1985. Based on the weight of overall evidence from human, animal, and mutagenicity studies, the Agency concludes, in the current draft document, that 1,3-butadiene is a known, human carcinogen.

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2.2 The Review and Charge

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On April 30 and May 1, 1998, the EHC Committee met at the EPA's Waterside Mall Complex in Washington, DC to review the Agency's draft *Health Risk Assessment of 1,3-Butadiene* document. The Committee was charged to provide comments on each of the following aspects of the document:

a) Review the health risk assessment for technical quality, comprehensiveness and clarity (Address each chapter, but with specific reference to Charges b, c, and d).

b) Does the science support the classification of "known" human carcinogen?

c) Are the approaches taken to characterize plausible cancer risks reasonable given the science?

d) Are the conclusions and quantitative estimations for reproductive/developmental effects adequately supported?

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3. RESPONSE TO THE CHARGE

This report captures comments and recommendations made in consensus by the Committee as well as specific technical comments and recommendations provided by individual Committee Members. It was not possible to review and achieve consensus for each and every technical point provided in this Committee report. On major points an attempt has been made to reflect the consensus or range of views on the Committee.

3.1 Technical Quality, Comprehensiveness and Clarity

The Environmental Health Committee was asked to: *review the health risk assessment for technical quality, comprehensiveness, and clarity.* The Committee provided comments and recommendations for each chapter.

3.1.1 Chapter 1 - Introduction

The Introduction presents a clear and comprehensive summary of information on 1,3-butadiene for the period from 1985 until January 31, 1997. It was concluded that apparent differences in assessments by different groups might be explained by the availability of studies at the time of evaluations, different cancer classification systems, and quantitative assessments done for different purposes. Does this mean that they are all equally valid and scientifically defensible? In other words can the statements made in Table 1-1 be restated in comparable terminology to show that they are all compatible? If they are, as implied, then the document should demonstrate this in subsequent chapters. If not, the Introduction should state so, and subsequent chapters should explain why. The EHC also recommends that concentrations be expressed in one constant unit throughout the document as well as in Table 1.

Every document must have a cut-off in time for completion. However, the International

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Agency for Research on Cancer (IARC, 1998) has recently re-evaluated the evidence on the carcinogenicity of 1,3-butadiene, and Health Canada is completing a health evaluation on the compound. It will be important to include the IARC evaluation in an updated version of the health assessment, and should the Health Canada document be finalized soon, it should also be included. For example, Table 1-1 should include the IARC evaluation, and if available, the finalized Health
Canada evaluation (IARC, 1998). Also, the Agency should clarify, in the Table, whether the
OSHA classification listed 1,3-butadiene as a "known" or "potential" carcinogen.
It is stated that the profile for setting the American Conference of Governmental Industrial
Hygienists-Threshold Limit Value (ACGIH-TLV) for 1,3-butadiene is not reviewed because it is
not a risk assessment. This may be true for the EPA definition of a risk assessment, but the
Agency should include some history of what has been used for the work environment, particularly
because of the high values in the past and the importance of epidemiological studies on worker
population.
There should be a comment in the final document on the accessibility of unpublished data
that is referenced in Agency documents. This concern was based on the use of the
Delzell/University of Alabama at Birmingham (UAB) study in the Agency's health risk assessment
of 1,3-butadiene even though the review document was not readily available to the public
(Delzell, 1995).
Finally, there is no mention in the Introduction of EPA's proposed Cancer Risk
Assessment Guidelines. While these guidelines have not been completely finalized, they are
sufficiently complete to be referenced and utilized as appropriate (USEPA, 1996).

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		3.1.2 Chapter 2 - Overview of Exposure to 1,3-Butadiene
	This	chapter is intended to be an introductory review of possible sources of exposure to
1,3 l	butadien	e. It should be clearly indicated in the first paragraph that this chapter is not intended
to be	e a comp	prehensive review of exposure so that this chapter is not mistakenly used as a source
of da	ata for e	nforcement purposes.
An e	explanat	ion should be included on how concentrations have been measured over the years and
how	this mig	ght impact a comparison of exposure levels measured in earlier years with more recen
mea	suremen	ts. Ranges of concentrations are important to include along with averages. The
Con	nmittee 1	recommends that all of the exposure concentrations be expressed in one common unit
thro	ughout (Chapter 2.
Add	litional p	oints that need to be incorporated include:
1.		
	a)	Section 2.2.1. Butadiene monomer production facilities need to be discussed
		especially since the epidemiology section discusses exposure in these facilities
2.	b)	Section 2.2.1. The impact of compliance with the 1994 Hazardous National
		Emissions Standard for Hazardous Air Pollutants and the polymers and resins
		Maximum Achievable Control Technology (MACT) on air emissions should be
		discussed.
3.	c)	Section 2.3.3. This section discusses open burning of tires. Data on controlled
		burning of tires should also be included and distinguished from open burning.
		Specifically, results from a pilot study conducted by Paul Lemieux (1994), US

EPA Air and Energy Engineering Research Laboratory should be

included. In

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throughout the document.

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1 this study, 1,3 butadiene could not be detected from controlled combustion of 2 tire-derived materials. 3 4 4. d) Section 2.4. For a better perspective, where possible, tables should indicate 5 the number of samples that were below the detection limit as well as the 6 ranges of exposure concentrations. More recent data on emissions, such as the 7 1995 and 1996 Toxics Release Inventory (TRI), and soon to be released 8 EPA's 1996 nationwide emissions inventory for butadiene should be included 9 (USEPA, 1997, 1998b). If available, exposure concentrations inside 10 automobiles should be added. 11 12 Section 2.5. In this discussion of exposures, similarities and differences of general e) 13 population and occupational cohort exposures should be discussed so that the 14 limitations of extrapolation from occupational setting to exposure of 15 population is understood. For example, short peak exposures in industrial setting 16 vs. chronic low levels of exposure to general population may impact extrapolation of occupational data. 17 18 19 3.1.3 Chapter 3 - Metabolism and Pharmacokinetics 20 21 Chapter 3 reviews the metabolism and pharmacokinetics of 1,3-butadiene. This chapter is 22 long, contains numerous incorrect or incomplete statements that need to be changed, and should 23 be updated. The title of the chapter should be Metabolism and Toxicokinetics, as butadiene is not

used as a therapeutic agent. Also, the word, toxicokinetics, should replace pharmacokinetics

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The Health Canada assessment of butadiene and Dr. Himmelstein's submitted comments will provide many of the new references that need to be incorporated into the revision. Specific issues that should be addressed in the revision of the risk assessment are provided below (Health Canada, 1997) (Himmelstein, 1998).

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In evaluating the data on the metabolism of butadiene and its epoxide metabolites across species, including humans, the EPA concluded that there are no clear species differences in butadiene metabolism and that in many cases there is overlap in metabolic rates across species. In forming their conclusion, the EPA misinterpreted the in vitro data and ignored the major conclusion from the in vivo data. With respect to the in vitro data, the EPA compared maximal rates of metabolism (i.e., V_{max}) rather than basing their comparisons on a more appropriate measure of metabolic rates, the ratio of V_{max}/K_m . This ratio is more appropriate because a) species differences are observed in both metabolic parameters, and b) at non-metabolically saturating concentrations (i.e., those concentrations relevant for humans) it is the ratio, not the maximum rate, that dictates the rate of metabolism. With respect to the in vivo data, the EPA has not adequately summarized the data from two independent laboratories that clearly and unambiguously show large differences spanning orders of magnitude in the blood and tissue concentrations of the butadiene metabolite diepoxybutane. In particular, Table 3-8 is misleading. This table includes data from the rat, mouse, and monkey for blood epoxybutene and diepoxybutane concentrations from studies conducted by Bond et al. (1986) and Dahl et al. (1991) which did not use gas chromatography-mass spectrometry (GC-MS) methods to quantitate these metabolites (Bond, et al., 1986)(Dahl, et al., 1991). Therefore, the values reported by these investigators are unreliable. It would be preferable to, instead, compare the blood concentrations reported by Himmelstein, et al., Bechtold et al., and Thorton-Manning et al. in one table (Himmelstein, et al., 1994, 1995))(Bechtold, et al., 1995)(Thorton-Manning et al., 1995a, 1995b, 1996). This comparison would serve two purposes. First, it would highlight the interlaboratory reproducibility in the values reported for these metabolites serving to increase the

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reliability of the data. Second, it would accentuate the dramatic species differences in the circulating levels of diepoxybutane, lending support to the hypothesis that this interspecies difference in metabolite formation underlies the differences in susceptibility observed in the chronic studies.

Mutagenicity studies strongly suggest that diepoxybutane is **a**, if not, **the** critical metabolite in butadiene carcinogenicity. Thus, differences in levels of this metabolite formed in vitro and in vivo are highly consistent with the observed species differences in carcinogenicity. The Committee recommends that the Agency integrate and assess this information, develop conclusions based on the weight of all the information presented, and present these conclusions in the closing paragraphs of Chapter 3.

It is recognized that the EPA elected a cutoff date for inclusion of new information of January 31, 1997. However, there is no question that a significant amount of new and important information has been developed since this date that is particularly pertinent to Chapter 3 on Metabolism and Pharmacokinetics. In short, the chapter on Metabolism and Pharmacokinetics needs to be updated to include key references that have been published in the peer-reviewed literature since January 31, 1997. Many of the data sets that have been published since this date lend further support to the hypothesis that diepoxybutane is a critical metabolite involved in the carcinogenicity of 1,3-butadiene.

While this chapter references important literature relative to the in vitro and in vivo metabolism and butadiene, it falls short in providing a comprehensive integration of the relatively diverse data sets. A critical concept that appears lacking in this chapter is the fact that diepoxybutane represents a critical metabolite of butadiene and that there are significant species differences both in vitro and in vivo in the formation of this important butadiene metabolite. This chapter neglects to note that metabolism and toxicokinetic studies of butadiene conducted in

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whole animals and in rodent and human tissues provide important insights into the likely critical steps in the initiation of butadiene carcinogenicity and importantly the identity of the most likely chemical species responsible for the development of tumors. For example, dosimetry data on both epoxybutene and diepoxybutane following inhalation exposure to butadiene clearly indicate that blood concentration of epoxybutene were up to 8-fold higher in mice compared with rats and that blood concentrations of diepoxybutane were nearly 40-fold higher in mice than in rats. Similarly, tissue concentrations of epoxybutene ranged from 3-10 times higher in mice compared with rats and tissue concentrations of diepoxybutane were up to 100 times higher in mice than rats. Importantly, therefore the correlation between measured circulating blood and tissue levels of the epoxides, particularly diepoxybutane and the observed development of tumors is clearly suggestive of a role of diepoxybutane in the initiation of cancer. This important concept is not presented in the chapter.

Moreover, in vitro data on the metabolism of butadiene suggest that mice form epoxybutene and diepoxybutane at a faster rate than rats or humans. Studies on the in vivo metabolism and tissue concentrations of epoxybutene and diepoxybutane in mice and rats following inhalation exposure to butadiene are consistent with the in vitro studies on metabolism of butadiene. This is an important point and the chapter neglects to point out the close parallels between the observations in in vitro studies and in vivo studies.

The Agency notes that butadiene is an animal carcinogen and that the mouse is more sensitive than the rat to butadiene induced carcinogenicity. The Agency also notes that the reasons for these interspecies differences are not understood at this time. However, the available mechanistic data on the metabolism of butadiene and its reactive epoxide metabolites supports the hypothesis that interspecies differences in metabolic rates form the underpinning for the increased sensitivity of the mouse compared with the rat. These differences in metabolic rates result in a faster production and slower detoxification of the diepoxybutane in mice compared with rats with

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resultant higher steady state levels of the diepoxybutane in blood and tissues in the mouse compared with the rat following exposure to butadiene. This observation coupled with the demonstrated hundred-fold greater mutagenicity of the diepoxybutane compared with the epoxybutene points to interspecies differences in the formation of diepoxybutane being critical to interpretation of differences in response.

The Agency notes on page 11-9 of the document that the specific mechanisms of 1,3-butadiene induced carcinogenesis are unknown. As noted above, there is extensive mechanistic data on the metabolism of butadiene in various species, including humans, that suggest that the extreme sensitivity of the mouse is related to the excessive formation of the diepoxide metabolite in this species. Experimental studies show that both humans and rats produce much less of the metabolite.

Additional points that should be incorporated include:

a) Page 3-1, Line 15: Much of the literature used the abbreviation BD for 1,3-butadiene. It should be less confusing if BDiol is used for the 3-butene-1,2-diol. This section does not even mention the 3,4-epoxy-1,2-butanediol (EBD). The latest research on molecular dosimetry strongly suggests that EBD is the major electrophile that binds to DNA and hemoglobin. EBD should be readdressed in the final version of this risk assessment.

b) Page 3-2, Line 14: Recent data shows that the trihydroxybutane (THB) adducts are clearly the predominate adducts in DNA and hemoglobin. These can arise from either 3,4-epoxy-1,2-butanediol or 1,2:3,4-diepoxybutane (DEB). Based on tissue measurements of DEB and EB, the ratios of epoxybutene (EB) to DEB/EBD(THB) adducts strongly suggest that EBD is the primary source of the

1		THB adducts. The EBD metabolite has not been quantitated following in vivo
2		exposure. This represents a major gap in our knowledge that needs to be
3		acknowledged.
4		
5	c)	Page 3-2, Line 14-18: The toxicokinetic data clearly does not support the
6		statement that the possible crotonaldehyde metabolites are "Of greater
7		significance." This editorial comment should be removed from the document, since
8		no causal role of these metabolites in butadiene mutagenicity or carcinogenicity has
9		been shown.
10		
11	d)	Page 3-3: This chart has several errors that need correction. 3-Butene-1,2-diol is
12		not reactive and should not have a box around it. Furthermore, the arrow for
13		Reaction 11 is going in the wrong direction. Crotonaldehyde and acrolien are
14		reactive and should have boxes around them. The urinary metabolites M-I and
15		M-II should be shown.
16		
17	e)	Page 3-27: (9) This reaction seems to be mislabeled. It appears that this refers to
18		BDiol GSH-BDiol.
19		
20	f)	Page 3-38, Line 21-22: Most 1,3-dihydroxypropanone probably comes from EBD.
21		This statement gives a wrong impression.
22		
23	g)	Page 3-39, Line 18-25: The toxicokinetics of butadiene are much more
24		complicated than discussed. The molecular dosimetry of DNA adducts following 4
25		weeks of exposure clearly shows that the first oxidation to EB and its subsequent
26		binding to DNA is linear over a range of 20-625 ppm in rats and mice (Swenberg,
27		et al., 1998). It is rapidly converted to BDiol and then to EBD. To a lesser extent,

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1 EB is oxidized to DEB. Both EBD and DEB form THB adducts, although most of 2 these come from EBD. The formation of THB adducts is saturated at 62.5 ppm in 3 the rat, so that exposure to higher amounts such as 1000-8000 ppm results in little 4 more THB adducts than does exposure to 62.5 ppm. In contrast, the mouse shows 5 a biphasic response for the formation of THB adducts, with a steep slope between 6 0 and 62.5 ppm and a lesser slope from 62.5-625 ppm. The mouse does not show total saturation of the formation of THB adducts, suggesting that it has a second 7 enzymatic pathway that is still active at high exposures. These data were presented 8 9 at the SAB meeting, were presented at the Society of Toxicology (SOT) and 10 Health Effects Institutes (HEI) annual meetings in 1998, and will be submitted for publication this summer (Tretyakova, et al., 1998)(Swenberg, et al., 1998). 11 12 13 h) Page 3-40, Line 13-14: This sentence is unclear. What is metabolic capacity of 14 EB? Is this formation or further metabolism? Available DNA data suggests that 15 formation continues to occur in an exposure related manner. 16 17 I) Page 3-41, Line 8: Butene diol is not a hydrolysis product of DEB. This should be 18 EB. 19 20 <u>j</u>) Page 3, Line 41-42: The sections on DNA and hemoglobin adducts are seriously 21 out of date, as this is a fast moving area of research. Data from several laboratories 22 have shown that the THB adducts are predominant over EB adducts. At high 23 concentrations of butadiene (625-1000 ppm), the ratio of THB/EB is 1.5-4 for rats 24 and 3-9 for mice. This drastically changes at nonsaturating exposures of 20 ppm 25 and 62.5 ppm, where rats have 27.5 and mice have 43-47 times more THB 26 adducts. The same finding has been demonstrated for human hemoglobin adducts. 27 THB-Valine adducts are formed about 40 times more frequently than EB-valine

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adducts. The EPA document only mentions EB adducts. A molecular epidemiology study of hemoglobin adducts was recently completed at the National Cancer Institute (NCI) in a Chinese butadiene worker study coordinated by Dr. Richard Hayes. Nearly all blood samples examined, whether from exposed or unexposed individuals, had measurable THB-Valine adducts. The number of adducts in unexposed individuals averaged ~40 pmol/g globin. There are similar data for U.S. research workers. With exposure to butadiene estimated to be 1-3.5 ppm, the number of adducts increased 2-3-fold ($R^2 = 0.33$). Of interest is the finding that glutathione S-transferase theta (GSTT1) genotype had no effect on THB-Valine adducts. Since the GSTT1 null genotype has clearly been associated with increased susceptibility to DEB-induced sister chromatid exchanges (SCEs) and is primarily located in erythrocytes, this observation supports EBD as the primary human electrophile forming THB-Valine adducts. k) Page 3-45: The section on Discussion and Conclusions should be revised. Lines 8-10 are oversimplified. Line 12 is wrong. The 1,3-butadiene epoxide should be

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k) *Page 3-45*: The section on Discussion and Conclusions should be revised. Lines 8-10 are oversimplified. Line 12 is wrong. The 1,3-butadiene epoxide should be butene diol. Lines 19-23 clearly do not reflect molecular dosimetry data and actually appear to be reversed. Line 27 is wrong. Butene diol is not toxic. EBD needs to be added to this section, as it is likely to be a major metabolite.

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This section of the risk assessment should also address what is known about genetic polymorphisms that are likely to affect individual susceptibility to butadiene and its metabolites. Several genes appear to be important. Inherent susceptibilities have been shown for both DEB and EB (Weincke and Kelsey, 1993), which may be due to glutathione S-transferase theta (GSTT1) status. Also, glutathione S-transferase $\mu(GSTM1)$ appears to be an important detoxifying factor for EB, so that GSTM1 null individuals would be expected to have greater effects following formation of EB. Unfortunately, no data have been published on the effects of GST

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polymorphisms on EBD. This is a gap in our knowledge. Genetic polymorphisms have also been
identified for epoxide hydrolase (EH) and CYP 2E1 that would be expected to affect
susceptibility to butadiene and its metabolites. The role of these proteins in the toxicokinetics of
numerous chemicals is reasonably well known. Three studies (Csanday, et al., 1992; Seaton et al.
1995, and Duescher and Elfarra, 1994) have shown in vitro using rodent and human tissue
samples that CYP2E1 plays a role in the oxidation of both BD and EB. It is possible to expect
that polymorphisms that reduce EH activity will increase susceptibility to butadiene. Likewise,
rapid CYP 2E1 metabolizers would be expected to be at greater risk.

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314	Chanter 4 -	- Mutagenicity

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This chapter summarizes the genotoxicity of 1,3 butadiene (BD) and its metabolites for mice, rats and humans, considering both cytogenetic endpoints and gene mutations in somatic and germinal cells. The emphasis is on recent in vivo studies, although there is some mention of results from in vitro assays. Most of the extensive work prior to 1994 is not mentioned. However, citations are given for the numerous relevant reviews. The recommendations for Chapter 4 are given below.

- a) Text tables summarizing key animal and human findings derived from the entire body of information on BD should be added.

b) References to support key findings should be included in the tables.

c) Separate tables should be included for in vitro, animal and human findings.

d) Similarities and species differences in response should be noted.

e) More emphasis should be given to the positive heritable translocation studies in mice because of their potential relevance for human heritable risks, and to several additional studies conducted in humans. This recommendation is also discussed in Section 3.1.5 which addresses the *Reproductive and Developmental Effects* chapter of the review document.

f) Some of the key findings that should be added or expanded are discussed in detail below. The conclusion section should be expanded to include what is known about the mutagenicity of BD and its metabolites from the extensive literature. Careful editing of the final document should be conducted to avoid missing dose units,

1		units for mutant frequencies, and similar omissions.
2		
3	g)	BD Metabolites Bind to DNA
4	i)	Electrophilicity is an important property of chemical mutagens. The
5		evidence that BD's metabolites react with proteins such as hemoglobin is presented
6		in Chapter 3 of this draft document. Evidence for reactivity with the DNA
7		itself demonstrates that these BD intermediates reach their target molecule for
8		genotoxicity causing premutagenic DNA lesions. Some of this evidence is
9		also reviewed in Chapter 3, but should be expanded greatly in this chapter
10		on mutagenicity.
11		
12	ii)	Early studies showed that DEB binds to N-7 guanine and that it forms inter-strand
13		crosslinks (Brooks and Lawley, 1961; Lawley and Brooks, 1967).
14		Subsequently it was shown that B6C3F1 mice and Wistar rats exposed to ¹⁴ C
15		labeled BD by inhalation have covalently bound reactivity in liver DNA, as
16		noted in Chapter 3 (Kreiling et al., 1986). The amounts bound in the two
17		species were comparable.
18		
19	iii)	The complex kinds of adducts formed in DNA by BD metabolites are also
20		under active investigation. Specific enantio- and regioisomeric EB adducts
21		formations have been shown (Koivisto et al., 1997; Tretyakova et al., 1997).
22		The N-7 position of guanine has been shown to be the most reactive with EB,
23		followed by the N-3 and N-1 positions of adenine. EB adducts have also been
24		found at the N-6 position of adenine but this may represent a rearrangement of
25		the N-1 adenine adduct. Adenine adducts may be important for the
26		genotoxicity of BD as shown by mutational spectral studies of this agent.
27		

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2	iv)	The several nucleobase adducts in DNA formed by BD metabolites are
3		being characterized further by several groups (Leuratti et al., 1994; Neagu
4		et al., 1994, 1995; Selzer and Elfarra, 1996; Kumar et al., 1996).
5		Furthermore, attempts are being made to use high sensitivity methods for
6		detecting DNA adducts in humans for biomonitoring purposes, including
7		the detection of urinary DNA adducts. However, despite the statement
8		made in Chapter 11, page 11-9, para. 1, line 2, the Committee is not
9		aware that DNA adducts have even been observed in vivo in humans. The
10		statement in this regard made on page 11-9 either needs a reference or
11		should be corrected.
12		
13	h)	Extensive Genotoxicity Profile for BD and its Metabolites
14		The extensive mutagenicity results covered in the several reviews cited in Chapter
15		4 include positive Salmonella assays in the presence of S9. It is important to note
16		that human S9 also converts BD to mutagenic metabolites, as determined in the
17		Salmonella system (Arce et al., 1990). BD metabolites also have been positive for
18		mutagenicity in a variety of microorganisms with or without metabolic activation.
19		Studies of mutation by BD at the tk locus in mouse lymphoma cells have been both
20		positive (Sernau et al., 1986) and negative (McGregor et al., 1991). BD, as
21		opposed to its metabolites, has not been found mutagenic in vivo in Drosophila
22		melanogaster, as assessed by the sex-linked recessive lethal mutation assay
23		(Foureman et al., 1994) or by the Wing Spot Test (Victorin et al., 1990).
24		
25	i)	Comparative Genotoxicity Results in Mice and Rats
26		There have been numerous cytogenetic studies in vivo in rodents. These are
27		covered in the cited reviews and, as correctly stated in Chapter 4, support the

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dichotomy in carcinogenic response between these two species, where mice are more responsive than rats. However, it should be noted that, with regard to cytogenetic assays in vivo in these two species, there are no reports of positive results in rats exposed to BD (as exposed to BD metabolites) but there are several reports of positive results in mice.

- j) Contrast in clastogenicity results between species
 - i) The difference in BD's apparent mutagenic potencies for mice and rats are worthy of comparison. Studies can be considered as those measuring clastogenicity and those measuring specific gene mutations. In regards to clastogenicity, for mice there are numerous reports of chromosome aberrations, micronuclei and SCEs in somatic cells, i.e. in both lymphocytes and in bone marrow cells, several showing effects at low doses. However, in rats, neither chromosome aberrations nor micronuclei have been found in blood cells in vivo (i.e. in lymphocytes, bone marrow or peripheral red blood cells) after BD inhalation (Arce et al., 1990; Autio et al., 1994), although there is a report of a weak positive SCE response (Maki-Paakkanen et al., 1993).

ii) For germ cells, the contrast in clastogenicity results between these species is equally striking. In mice, there are reports of dominant lethal effects with inconsistent results (references given in the draft document), sperm head abnormalities (Morrissey et al., 1990), micronuclei in spermatids (Xiao and Tates, 1995; Tommasi et al., 1998) and a positive Comet assay in haploid and polyploid testicular cells (Brinkworth et al., 1998). The dominant lethal studies discussed in Chapter 5 should be cross-referenced in Chapter 4 as they are important to the discussion of heritable chromosomal alterations

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1 induced by BD. Cytogenetic abnormalities in first-cleavage embryos sired 2 by male mice treated by BD inhalation have also been reported recently 3 (Pacchierotti et al., 1998a). By contrast, in the single study in rats, no 4 dominant lethal mutations were found (BIBRA, 1996). 5 6 iii) Of special importance among the studies of clastogenicity are those of 7 heritable translocations in mice. The first study is discussed briefly in 8 Chapter 4 (Adler et al., 1995). A second study by the same group has 9 recently been reported (Adler et al., 1998). These studies are particularly 10 relevant to human health and risk assessments for human heritable damage 11 and should be emphasized. In fact, a human risk estimate has recently been 12 reported using a parallelogram approach that employed mouse somatic cell 13 clastogenicity (micronuclei in bone marrow cells), mouse germ cell 14 clastogenicity (the heritable translocations) and human somatic cell 15 clastogenicity (in lymphocytes from BD exposed workers) to estimate the heritable translocation risk for humans (Pacchierotti et al., 1998b). The 16 17 estimated doubling dose for human heritable translocations was given as 18 1,100 ppmh (parts per million hours) 19 20 Comparison of the specific locus mutagenicity of BD between mice and rats is not k) 21 nearly as striking as is the comparison for clastogenicity. The studies summarized 22 by Recio and Goldsworthy (1995) showing an increase in BD-induced lacI mutations at A:T base pairs in bone marrow stem cells from young transgenic 23 24 B6C3F1 (BB) male mice are discussed in Chapter 4. However, it should be 25 mentioned that an earlier study in young male Balb/c X DBA/2 (CD2F1) 26 transgenic mice (MM) found lacZ BD induced mutations only in lung cells and not 27 in liver or bone marrow stem cells (Recio et al., 1992). There is also a report of a

1		positive spot test in "T" stock mice that found an increase in in vivo mutations in
2		embryonic melanoblasts (Adler et al., 1994).
3		
4	1)	There are several reports of positive hprt mutations in young B3C3F1 mice
5		exposed to BD by inhalation. The Cochrane and Skopek (1994b) study
6		mentioned in Chapter 4 exposed pre-weanling mice and found dose-related
7		mutational increases. Again, there was a bias for A:T changes. The Meng study
8		mentioned in Chapter 4 (Meng, et.al, 1996) also exposed young B6C3F1 male
9		mice to BD and found increases in hprt mutations in both thymic and splenic
10		lymphocytes that persisted for several weeks after the exposures. This work has
11		now been accepted as a full publication and indicates that the hprt mutant
12		frequency (MF) in thymic lymphocytes rises from 2.2×10^{-6} to 11.3×10^{-6} at two
13		weeks (maximal) and, in splenic lymphocytes, from $\sim 1.8 \times 10^{-6}$ to 19.7×10^{-6} at
14		five weeks (maximal). The mutagenic potency of BD in these mice, which is a
15		measure of hprt mutant cell accumulation, was calculated to be 69.62.
16		
17	m)	There are also negative reports for in vivo hprt mutations in other strains of mice
18		that were older at the time of their BD exposures. Tates et al. originally found low
19		order hprt mutagenicity in splenic lymphocytes of 10-12-week-old (102/E1 X
20		C3H/E1) F1 mice (Tates et al., 1994), but later failed to find an increase in such
21		hprt mutations in a second study of this strain or in adult CD1 mice (Tates et al.,
22		1998).
23		
24	n)	In contrast to the clastogenicity studies, specific locus gene mutations have been
25		found in vivo in rats exposed to BD by inhalation. Meng et al., 1996 exposed
26		young rats with results as noted in Chapter 4, i.e. increases in hprt mutations in
27		thymic lymphocytes from 2 x 10^{-6} to 4.9 x 10^{-6} at three weeks after exposure

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(maximal) and, in splenic T-cells, from ~1.9 x 10⁻⁶ to 10.1 x 10⁻⁶ at four weeks 1 2 after exposure (maximal) (references given in chapter). The mutagenic potency of 3 BD in rats was calculated to be 15.85. Thus, the ratio of mutagenic potencies between mice and rats (mice/rats) was calculated to be 4.4 or 5.0, depending on 4 5 the weeks allowed for mutant cell accumulation. Thus, although BD is 5X more 6 mutagenic in rats, the magnitude of this difference is not nearly as great as that 7 reported for the carcinogenicity differential between these two species. 8 9 0) Studies of BD Metabolites 10 The many rodent studies of BD metabolites, some positive and some negative, 11 show a similar pattern. In general, younger animals tend to give the positive results 12 and, for mice, it is the B6C3F1 animals that are most often associated with positive 13 studies. The Cochrane and Skopek studies of EB and DEB were noted in Chapter 14 4. Meng et al. have extended their studies in young mice (B6C3F1) and rats 15 (Fischer) to EB and DEB by inhalation and have shown increased hprt mutations 16 in splenic TÄcells in both species (Meng et al., 1997). In rats, however, the 17 response to EB was equivocal. In contrast to these positive results, Tates et al. 18 found no hprt mutation induction in older mice (102/E1 X C3H/E1) F1 and CD1 19 or rats (Lewis) administered EB or DEB by injection or in the drinking water. 20 However, as noted in Chapter 4, rat germinal cells are at least as susceptible (or 21 more so) to the clastogenicity of BD metabolites as are mouse germinal cells. 22 23 Summary of In Vivo Rodent Studies p) 24 i) In summary, there are striking species differences between mice and rats in 25 the reported studies as regards clastogenicity. There are numerous positive

studies of this endpoint in mice, but BD induced clastogenicity has not been

demonstrated in vivo in rats. Of note, heritable translocations have been

1		demonstrated and confirmed following BD exposures to mice.
2		Clastogenicity has been observed in mice and rats exposed to EB or DEB.
3		
4	ii)	For gene mutations, B6C3F1 mice appear to be more sensitive than other mouse
5		strains to the mutagenic effects of BD, EB or DEB. In general, mice are
6		also more sensitive than rats to specific gene mutations, although such
7		mutations can be induced in younger rats. Mutations are most commonly
8		seen in both species in young animals, indicating that cell proliferation may be
9		required to produce these mutagenic effects.
10		
11	q)	Human Studies
12		i) Chapter 4 includes most of the relevant mutagenicity (clastogenicity and
13		specific gene mutations) studies in human cells and/or in vivo in humans.
14		
15		The Cochrane and Skopek study (1994) of hprt and tk mutations in vitro in
16		human TK6 cells described in page 4-1 should note that mutation induction
17		was assessed for EB, DEB and for EBD. The mutagenic potencies of
18		these BD metabolites were found to be DEB> EB>EBD. It is important to
19		note that the metabolite EB diol has been studied and has been found to be
20		mutagenic. The importance of EBdiol lies in its abundance. Therefore, even
21		though Ebdiol(EBD) may be the least mutagenic of the metabolites, it may
22		be the most abundant, and therefore may give the most mutations.
23		
24	ii)	Some of the in vivo cytogenetic studies in humans are covered in Chapter
25		4. The positive challenge assay reported by Au et al., 1995 is noted as is
26		the negative report of chromosome aberrations, micronuclei and SCEs in
27		exposed workers by Sorsa. Not noted, however, is the update by Sorsa

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1 (Sorsa et al., 1996) that reanalyzed these data according to GST T1 status 2 and found that the T1 null workers appeared to have increases in chromosome 3 aberrations. Also, the increases in chromosome aberrations but not in 4 frequency of micronuclei reported by Tates in 1996 (and noted in Chapter 5 4) has been updated in a full manuscript that reports increases in both 6 chromosome aberrations and SCEs in BD exposed Czech workers. The 7 frequencies of micronuclei were not increased and the Comet assay was 8 negative (Tates et al., 1998). 9 10 iii) The in vitro findings of increased EB induced SCEs in lymphocytes from 11 GST M null individuals and increased DEB induced SCEs in lymphocytes 12 from GST T null individuals are reported in Chapter 4 and are important, 13 as is the observation that GST status does not appear to affect SCE 14 frequencies induced by EBD. A recent in vitro study observed chromosome 15 specific aneuploidy (for chromosomes 12 and X) in human lymphocytes in 16 vitro following treatments with either EB or DEB (Xi et al., 1997). The 17 paper by Wiencke et al, (1995) demonstrating the affinity of the glutathione 18 transferase theta enzyme for the diepoxide metabolite of BD should be 19 added. 20 21 iv) The status of the in vivo hprt mutation studies in humans has been 22 accurately reported in Chapter 4. There are the three positive studies using 23 the autoradiographic assay, as noted, and the two negative studies that 24 employed the cloning assay. Chapter 4 discusses the discordance in these 25 two groups of studies and concludes that, regardless of the reason for the

difference, the positive results with the autoradiographic assay probably

reflect a mutagenic effect of BD in the monitored workers. . There was a

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difference in opinion among the Committee regarding the hprt results. Some of the Committee found the Agency's conclusions to be reasonable while other Members/Consultants were concerned that the autoradiographic results may be method-related.

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The explanation as to possible reasons for the discordance between the positive effects obtained by autoradiography and the negative results found by cloning for human BD induced hprt mutations in vivo however should be rewritten. In considering the differences between the Ward et al., 1994, 1996 positive autoradiographic studies and the Hayes et al., 1996 and Tates et al., 1996 negative cloning assay studies, the draft document concludes that "a simple explanation would be that the increase in the autoradiographic assay was due to clones of mutants having arisen from earlier mutations." There was a diversity of opinion regarding the discordance between the positive effects obtained by the autoradiography and the negative results found by cloning for human butadiene inducted hprt mutations in vivo. Some of the Committee found that Agency's explanation is probably not correct for two reasons. First, the autoradiographic assay requires a technical step before the T-cells can be assessed for hprt mutations, i.e. cryopreservation or some measure to arrest those few cell that are cycling in human peripheral blood at the G1-S interphase. This is simply a technical means to insure that cycling nonmutant T-cells do not progress to their S-phase in vitro in the presence of 6-thioguanine and become labeled - even slightly labeled - and thereby become scored as mutants in the assay. Such cells are not mutants but would appear to be so by virtue of their (partial?) labeling, i.e. they are phenocopies. This phenomenon, which artifactually elevates scored variant

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1 frequencies, can be eliminated by arresting the cycling cells at this phase of 2 the cell cycle, from which they rapidly proceed into S in vitro before the 3 label is added. Thus, they miss the scoring window and are not scored as 4 variant cells. 5 6 vi) The reason why this arresting or cryopreservation step is relevant to the 7 issue of clonality is that T-cells that are undergoing clonal expansions in 8 vivo tend to be among the cycling cells. If such large mutant clones were 9 present, the cycling members of the clones would be eliminated from the 10 scoring window by the cryopreservation step (even though these would be 11 mutants). Therefore, if anything, clonal amplifications are less likely in the 12 autoradiographic than in the cloning assay. Said the other way around, if in 13 vivo clonality were the reason for the higher mutant frequencies in exposed 14 vs. control workers, this phenomenon would have most likely affected the 15 results in the cloning assay. This is the exact opposite to what was 16 observed, i.e. the increased mutant frequencies in exposed over controls 17 were observed with the autoradiographic assay. 18 19 Another reason why in vivo clonality cannot account for the differences in vii) 20 results between the assays is that molecular studies were not done. Such 21 studies cannot be done for the autoradiographic assay but can when using 22 the cloning assay. Molecular analyses allows detection of in vivo clonality 23 using the cloning assay. Since no such studies were done, no "corrections" 24 were made and the point is moot. 25 26 viii) There are other differences between the autoradiographic and cloning

assays that could account for the differences in results obtained with these

1		two assays. The autoradiographic method is a phenotypic assay, meaning
2		that mutants (or variants) are consumed by the assay and, therefore, cannot
3		be analyzed at the molecular level. It is not, therefore, possible to
4		demonstrate that the observed mutants scored by this assay are actually
5		genotypic mutants. This, however, is not different from other phenotypic
6		assays for mutations that are in wide use. To be scored in the
7		autoradiographic assay, variant cells have simply to synthesize DNA in
8		their first S in vitro. No cell division or growth is required. In addition, the
9		cells must begin their DNA synthesis within 36-48 hours in culture.
10		Therefore, a quite different T-cell subpopulation may be scored in the
11		autoradiographic than in the cloning assay. It is possible that, in the former,
12		only that subset of cells that are capable of division quickly in vitro is
13		scored whereas, in the cloning assay, all T-cells are scored. Alternatively,
14		what is being scored in the autoradiographic assay may not be fixed
15		mutations but rather adduct-blocked RNA transcription. Another
16		possibility is that the cell that would go into a G2 block, and therefore not
17		be measured by the cloning assay, are scored in the autoradiographic assay
18		because they are not required to go through G2 for scoring.
19		
20	ix)	The autoradiographic assay requires that peripheral blood T-cells be
21		cryopreserved in order to arrest cycling cells at the G1 S interphase. If all
22		cycling cells are not eliminated, the autoradiographic assay might give an
23		artifactually elevated reading because of the scoring of phenocopies. This is
24		explained above.
25		
26	x)	As for all mutation assays, the autoradiographic assay may have a scoring
27		bias. However, the cloning assay too has an inherent observer bias and a

1		tendency of some technicians to score only larger mutant colonies.
2		
3	xi)	In addition, with respect to the BD studies, the differences between the
4		positive and negative studies may be due, in part, to differences in study
5		populations. The Ward et al. 1994, 1996 radiographic studies were
6		conducted at a butadiene monomer production facility and an SBR
7		production facility in Texas. The Tates et al., 1996 study was conducted in
8		the Czech Republic (butadiene monomer production) and the Hayes et al.,
9		1996 study in China (polybutadiene rubber production). Smoking and other
10		lifestyle and confounding parameters would be different among these
11		populations. In addition, the exposure assessment in these studies were not
12		precise and no attempt was made to determine mutation susceptibility
13		profiles of the monitored workers. (It is of note, however, that one of the
14		highest variant frequency values in the Ward et al., 1996 study was in a
15		GST T null individual.)
16		
17	r)	Conclusions
18		Few conclusions are made at the end of Chapter 4. The quality of the conclusions
19		would be improved by expanding this section and adding statements summarizing
20		what we know about the mutagencity of BD and its metabolites. The following list
21		could serve as a guide:
22		
23		i) Rodent studies indicate that some of the metabolites of BD are DNA binding
24		agents.
25		
26	ii)	There are numerous in vitro assays in several model systems that
27		demonstrate the genotoxicity of BD and/or its metabolites.

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2	iii)	In vivo studies in mice and rats show that the former is the more sensitive
3		species for the genotoxicity that follows exposure to the parent BD.
4		
5	iv)	Young animals (including mice), and perhaps certain strains within a
6		species, are more susceptible to the genotoxicity of BD and/or its
7		metabolites than are others as indicated by hprt results.
8		
9	v)	Clastogenicity has not been demonstrated for either somatic or germinal
10		cells in rats exposed to the parent BD.
11		
12	vi)	Heritable translocations are induced by exposure of male mice to BD by
13		inhalation. (This is an important point for estimating heritable risks.)
14		Equivalent data are not available in the rat. Any estimation of human
15		heritable risk must account for species differences in metabolism.
16		
17	vii)	BD metabolites give genotoxic effects in both rats and mice with mice
18		being the more sensitive to somatic effects but both species being equally
19		susceptible (or rats more so) to the germinal effects.
20		
21	viii)	Human studies have shown that BD metabolites cause gene mutations in
22		vitro in human cells.
23		
24	ix)	GST genotypes may affect susceptibility to the clastogenicity (and possibly
25		mutagenicity) of BD metabolites.
26		
27	x)	There is evidence of clastogenicity in vivo in human lymphocytes from

1		exposed workers.
2		
3	xi)	Hprt mutations have been shown in vivo in human T-lymphocytes of
4		exposed workers as documented by the autoradiographic but not by the
5		cloning assay for these events.
6		
7	3.1.5	Chapter 5 - Reproductive and Developmental Effects
8	It was	s evident to the Committee that the EPA had a large amount of material to review
9	since that the	re were over 20 animal bioassays on this topic area. In addition, many new studies
10	have come in	after the cut-off date for the assessment and, as noted below, this research will need
11	to be included	d in the report.
12		
13	a)	Organization
14		Because of the volume and the complexity of the studies and reports for
15		both endpoints, additional summary tables are necessary in Chapter 5. All
16		pertinent studies that are discussed in this chapter should be introduced in
17		a summary table at the start of each section for developmental and
18		reproductive toxicity. A good example is Table 5-13 which was prepared
19		for summarizing the structure-activity information.
20		
21	b)	Inclusion of negative data
22		Chapter 5 needs to include both positive and negative studies in the
23		assessment. For example, text for Table 5-13 says that "since no
24		non-neoplastic lesions were seen, then they were not included in the
25		table." The report should include positive and negative data so the
26		reviewer can develop a comprehensive understanding of the data base
27		supporting the assessment.

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c) Inclusion of new data

The health assessment needs to include more current research. Every effort should be taken to extend the time for data inclusion to as close as possible to the release date of the assessment. For example, because of the cut-off date, several new dominant lethal studies were not included. These 2 new negative studies need to be included in the reassessment. The BIBRA Study III (Brinkworth, M., et al., 1998), which was a repeat of BIBRA I (BIBRA International, 1995), was negative and needs to be considered in this health assessment especially since the BIBRA I study was used in one of the quantitative assessments (BIBRA III) (BIBRA International, 1996)(Brinkworth, M., et al, 1998). Written testimony from M. Christian identifying and criticizing these studies should be considered in the report (Christian, M., 1998).

It was difficult for reviewers of this health assessment to integrate

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d) Integration of Chapter 5 with other chapters

i)

18 the toxicological findings. A much greater emphasis on integrating 19 findings across the chapters is needed. For example, information 20 on toxicokinetics and metabolism needs to be fully integrated into 21 this chapter. The information on toxicokinetics and metabolism in 22 the review document appear to be almost an after thought. 23 Interesting data on specific ovotoxic metabolites are available, and 24 should be incorporated. The potential impact of including the 25 PBPK model in the assessment of reproductive and developmental 26 toxicity needs to be discussed. Does this help to explain dose 27 response differences across species for ovotoxicity? The Agency

- DO NOT CITE OR QUOTE -1 should explain this in the assessment. It is important not only to 2 show data but to discuss the implication of the data. 3 4 ii) Another example where integration is needed is with the general 5 toxicity chapter with Chapter 5. Was there any evidence for 6 neurotoxicity from general toxicity testing to suggest the need for 7 developmental neurotoxicity assessment? What about effects in 8 utero on ovarian or testicular development? The heritable 9 translocation data from Chapter 3 should be integrated with 10 Chapter 5. 11 12 iii) Because of the potential interrelatedness of the reproductive tumors 13 and reproductive organ atrophy data, the reproductive chapter 14 should have some overlap with the chapter discussing tumor results 15 for these reproductive organs. Also, Chapter 5 should include a 16 discussion on the mechanisms responsible for atrophy and 17 reproductive tumors. 18 19 Biological significance of atrophy/dominant lethals e) 20 This chapter should provide discussion on the biological significance of 21 ovarian and testicular atrophy. This is especially true as these endpoints 22 are modeled in Chapter 9. How are exposed rodents different than control 23 rodents? The Agency should explain its rationale for looking at ovarian 24 atrophy in aged rodents. 25 26 f) Additional information 27 The following issues should also be addressed in Chapter 5.

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DRAFT #4

1		i) The reader needs to understand the rationale for modeling the
2		dominant lethal data. The Agency should provide discussion that
3		supports the use of these endpoints in Chapter 9. Since new studies
4		on heritable translocations are available, these findings should be
5		integrated with the chapter on mutagenicity. This will provide
6		stronger evidence for the potential of this compound or its
7		metabolites to produce heritable and multi-generational impact.
8		The EPA should integrate the data so that the total picture of
9		research supports your identification of critical endpoints and
10		quantitative modeling.
11		
12	g)	Uncertainty
13		For both the reproductive and developmental endpoints, it is necessary to
14		list all assumptions, identify where the assessor is uncertain and identify
15		agency action taken to respond to that uncertainty. The Agency should
16		explain what the critical data needs are that will address this uncertainty
17		and should be as specific and explicit as possible.
18		
19	h)	Use of speculative and editorial type of sentences
20		The Agency should avoid the use of speculative and editorial types of
21		sentences. A specific example is on Page 5-28, lines 32-33, the assessment
22		speculates that the compound does not cross the blood-testis barrier. Why
23		does the Agency think that this is true? Is this consistent with the
24		chemistry of the compound (i.e. solubility)? Is it consistent with the
25		postulated effects of this compound on spermatozoa?
26		
27	i)	Recommendations regarding specific statements include:

DRAFT #4 - DRAFT DOCUMENT-- DO NOT CITE OR QUOTE -1 Pages 5-1, lines 29-31 i) 2 Reword this sentence to make it neutral. "Because the results were 3 not analyzed statistically and other details regarding the duration of 4 the mating periods were not present, it is not possible to conclude 5 that 1,3-butadiene either had or did not have an effect on fertility in rats." 6 7 8 Pages 5-9, lines 27-29 ii) 9 Under what condition is 108± ppm 4 vinyl-1-cyclohexene 10 identified? What is the stock solution? The missing information 11 must be added. 12 13 Pages 5-25, tables 5-13 iii) 14 Why are only some of the studies of structurally related 15 compounds listed on this table? Why are negative observations not included? The explanation given on pp.5-24, lines 17-19 was weak. 16 17 Since neoplastic lesions in reproductive organs were seen, these 18 findings should be added here, albeit identified as neoplastic or non 19 neoplastic. The title of this table does not limit effects on neoplastic 20 status. However, the table should not just give positive studies but 21 also include negative observations, like those in rats. The 22 observations of decreased Graafian follicules should be included if 23 significant (note significance status was not given in lines 25-27). 24

Pages 5-28, lines 15-17

iv)

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The summary introduces new structure-activity information from

Maronpot, 1987. This data should be introduced and integrated

- DO NOT CITE OR QUOTE -1 with Chapter 5 prior to the summary. See also summary lines 7-33. 2 3 4 *Pages 5-28* v) 5 The summary speculates about reduced steroidogenesis, has any 6 study measured steroid levels? 7 8 Pages 5-28, lines 32-33 vi) 9 The Committee did not see any data presented to support this 10 speculation, in fact the observed effects of 1,3-butadiene on 11 spermatozoa and spermatids (discussed in the next paragraph) 12 argues against this "protective" effect of this barrier. The 13 Committee recommends the removal of this sentence. 14 15 vii) Table 5-1 Table 5-1 has numbers in parenthesis that are not defined. The 16 17 numbers that are in the parenthesis need to be listed in the footnote. 18 19 Section 5.1.6 viii) 20 Sections such as this would be clearer if a data table would 21 accompany the text. For example, see Tables 5-4 through 5-12. 22 23 ix) Page 5-28, lines 11-17 24 The paragraph on ovarian lesions fails to present the well 25 established mechanism for ovarian toxicity and carcinogenicity. 26 DEB is highly toxic to the ovary, which makes it non-responsive to 27 FSH. Continual elevation of FSH results in the carcinogenicity.

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In summary, Chapter 5 does not prepare the reader for the discussion in Chapter 9 on the quantitative risk assessment for 1,3-butadiene. For example, how does the biology information affect your choice of models? In Chapter 5, the Agency briefly makes a statement about thresholds, but does not explain implications for quantitative risk assessment. How did the Agency decide to drop the highest dose levels, was this because of excessive toxicity? If so, the EPA should explain the rationale. What criteria will the EPA apply to let he reader know when the Agency will drop the doses?

3.1.6 Chapter 6 - Toxicity in Animals

This chapter provides detailed information about subchronic, chronic and carcinogenicity studies published from 1985 to present. Only three studies are mentioned in the subchronic section. It is unclear why other repeat dose studies are not reviewed. Have they **all** been incorporated in other chapters? Repeat dose *in vivo* mammalian studies of 1,3 butadiene would be appropriate for inclusion in this section unless it is explicitly stated that they are covered elsewhere in the document. It is also important to insure that the data from *in vivo* repeat dose studies get thorough review because the Inhalation Reference Concentration (RfC) value should be derived on the most sensitive adverse endpoint that is meaningful for human health. The most sensitive non-cancer biological effect of 1,3 butadiene currently cannot be determined from the health risk assessment document, as comparative assessments are not made in any of the chapters.

In the chronic and carcinogenicity sections, the use of frequent text tables helps to simplify and to provide clarity to the presentation. Since the chronic National Toxicology Program (NTP) bioassay is a part of EPA's quantitative risk assessment, the detailed coverage is warranted. The NTP study contains both chronic toxicity and carcinogenicity data and the study is discussed in both areas. In order to make the chapter more understandable, the Committee recommends that

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the Agency present the study details presented in the first section (chronic) and then refer to them in the carcinogenicity section rather than repeat the material in slightly different words. As it reads now, the review document gives the impression that they are different studies. The Agency should also use the same categories of tumors for presenting data from the continuous treatment, 9 and 15-month interim sacrifices and stop-exposure study in order to permit comparisons. Most of the time this was done. However, for target lymphatic tumors the categories are different, preventing a direct comparison.

A table summarizing the positive oncogenic findings across all studies specifying the dose tested, and the type of tumor that was significantly elevated, should be added because of the numerous organs and tumor types involved. Data from the rat oncogenicity study should be included as this information is important and is only indirectly mentioned in the risk assessment.

For clarity, it would be preferable to separate the carcinogenic evidence on the mammalian metabolites of 1,3 butadiene from the data on related chemicals and place it in a subsection by itself. The section on related chemicals should be updated to include the results from studies on styrene, isoprene and any other relevant chemicals.

At the meeting, a member of the public stated that the data in Table 6-8 is wrong and that "Poly 3" data should be used. The Committee feels strongly that the rat carcinogenicity data need to be presented in similar detail to the NTP data. In doing so, the reader would not have to find the old risk assessment to see the data.

The quality of the Discussion and Conclusion section is good. The observation that concentration, not time is a critical determinant of potency is interesting but it is not supported by a comparison of the tumor data presented in Table 6-4 and Table 6-8 from the NTP study (1993) for continuous lifetime treatment and stop-exposure study. It is rare that carcinogenic data are available from continuous, interim sacrifice and stop-exposure studies. Further analysis of a

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comparison of the data from continuous and lifetime treatment would be interesting to see if there is supporting evidence for a biological model of dose and time that could be used in the risk assessment.

3.1.7 Chapter 7 - Epidemiologic Studies of Carcinogenicity

It is unclear why this chapter presented details of all the historical reports for a given butadiene (BD) exposed group, since more recent reports supersede the previous ones by virtue of having longer follow-up and more numerous deaths. Chapter 7 should be restructured so that it is a detailed report on the latest follow-up of each epidemiologic study, with perhaps any additional analyses from previous reports that were not duplicated in the most recent report. More specific recommendations for Chapter 7 follow.

a) Discussion of butadiene exposure and lymphosarcoma/reticulosarcoma

The case for an association of butadiene exposure with
lymphosarcoma/reticulosarcoma (ICD 200, a no-longer-used subset of
non-Hodgkins lymphoma; hereafter called just "lymphosarcoma") is based on two
studies of butadiene monomer plants which the EPA draft report indicated had
excesses of lymphosarcoma. However, many of the findings did not support, or
called into question, the purported association, specifically:

i) Texaco Study

The largest study group was most recently reported by Divine and Hartman, 1996 and consisted of 2,795 workers with an average of 32 years of follow-up (Divine and Hartman, 1996). The overall observed/expected ratio (O/E) for lymphosarcoma was 9/4.7= 1.91 (95% CI= 0.87-3.6) which was not statistically significant. The SMRs for those employed <5, 5-19 and 20+ years were 2.6, 1.8 and 0.8 respectively, which is largely counter to expectations. The lymphosarcomas were concentrated among those first employed during World War II (WWII)(O/E= 7/2.9= 2.4, 95%

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Confidence Interval (CI) = $1.0-5.0$). One feature that suggests it might be
a real effect is that the WW II excess was limited to those in the group with
jobs that entailed higher exposures ("varied exposure" group). However,
this elevated risk among WW II workers showed an inverse association
with length of employment, in spite of the fact that the authors indicate that
high exposures still continued into the 1950's and 1960's which does not
lend plausibility to the association. Although there was an overall excess of
lymphosarcoma in the group with jobs that entailed higher exposures
(O/E=7/2.8=2.5, CI=1.0-5.1) in the Divine-Hartman study, the subset of
this group employed for $10+$ years showed no excess (O/E= $1/1.0$). Divine
and Hartman created an index of cumulative butadiene exposure based on a
job exposure matrix that considered job class and calendar time. Using a
time-dependent Cox regression model for cumulative exposure vs.
lymphosarcoma, there was not even suggestive evidence for an association
between exposure and risk (Relative Risk (RR)= 1.00, 95% CI= 0.97,
1.04). On Page 7-32, Line 6 (and also Page 11-6, Line 22), in order to
present a balanced summary of findings on lymphosarcoma and monomer
production, it should be stated here that there was no indication of an
exposure-response association, based on the latest follow-up by Divine and
Hartman (Divine and Hartman, 1996). Two other methods of analysis
also reinforced the null association. Regression analyses that modeled the
time spent in each of their six job classes as predictors were also
conducted; again, there was no indication of an association. In summary,
the case for an association between butadiene exposure and
lymphosarcoma in this study is weak.

The EPA report of this study either fails to note or at least to consider the implications of a number of the findings noted above that go against

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the likelihood of a causal association between butadiene and lymphosarcoma. In the chapter summary (Pages 7-31 - 7-32) the Agency cites positive results from earlier follow-ups of this cohort and one nominally positive result from this report as their summary of the study, without noting the important findings in the study that are not supportive of a positive association.

ii) Union Carbide Study

Ward et al., 1996 studied 364 employees who worked at one of three units that had produced butadiene monomer (Ward, et al., 1996). For lymphosarcoma the ratio of O/E= 4/0.69= 5.77 (95% CI= 1.6-14.8). These four cases had worked at a butadiene unit for 0.8, 2.9, 3.3 and 8.0 years, so the worktime of three of the four was relatively brief. Two of the four, with butadiene exposures of 2.9 years and 3.3 years, had worked in an acetaldehyde unit for 8 years and 29 years, and all four had exposure to a variety of other chemicals at the facilities. The fact that two cases had long-time exposure to acetaldehyde raises the possibility that those cases may have been associated with acetaldehyde rather than butadiene, and this possible confounder weakens the finding. The nature and extent of the

possible confounding by acetaldehyde are glossed over in the EPA 1,3-

iii) Shell Study

butadiene report.

A third study of 614 employees at a butadiene monomer production plant yielded negative results (Cowles, et.al, 1994). They were followed for an average of 12 years after entry to the study (which, because of the unusual criteria for study entry, was equivalent to 14-17 years of follow-up after first exposure) and had an average of 7.6 years of butadiene exposure.

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1		There were no deaths from lymphatic or nematopoletic cancer (1.2
2		expected). This fact was not even mentioned in the EPA report (Page
3		7-8). This cohort provides no support for an association between
4		butadiene exposure and lymphosarcoma, although the fairly short
5		follow-up time and relatively small sample size means the negative results
6		should not receive a heavy weight. Nevertheless, the EPA statement that
7		"this study failed to provide any negative evidence towards the causal
8		association" (Page 7-32, Lines 15-16; also Page 7-9, Line 22-23 and Page
9		11-7, Lines 6-8) seems to be an overstatement; the study does provide
10		some, albeit not compelling, negative evidence. In fact, it is at least
11		equivalent in size to the Ward et al study that is highlighted as providing
12		positive evidence (Ward, et.al, 1996).
13		
14	iv)	Delzell Study
15		A large study by Delzell et al. (1996) of 15,649 styrene-butadiene rubber
16		(SBR) workers found no association between butadiene exposure and
17		lymphosarcoma (Delzell, et al., 1996). Overall the lymphosarcoma O/E=
18		11/13.8 = 0.8 (95% CI= 0.4-1.4). In those with the greatest exposure and
19		latency (10+ years exposure and 20+ years since hire) there was no
20		elevation in lymphosarcoma risk (O/E= 4/3.9= 1.0, CI= 0.3-2.6). The
21		other SBR studies are essentially earlier versions of this study, with minor
22		differences in the cohorts, so this study summarizes the lymphosarcoma
23		results for the SBR data.
24		
25		It is also of note that the coding of the ICD 200 category
26		"lymphosarcoma and reticulosarcoma," as distinct from the ICD 202

category of other non-Hodgkins lymphomas (NHL) for which no

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butadiene-related excesses were found in the various studies, is very unreliable. In a review of medical records for death certificates coded as lymphomas (ICD 200 or 202), Matanoski et al., 1993 found that two were not lymphomas at all "and the other 10 were so poorly classified into the 200 and 202 codes on the death certificates as compared with the hospital records that we combined these ICD categories" (Page 369) (Matanoski, 1993). This calls into question the meaningfulness of the positive results reported above which were based on death certificate diagnoses of lymphosarcoma. Had the other categories of NHL (i.e., ICD 202) been put with this category, it is unclear whether there would be any excesses of NHL.

b) Discussion of butadiene exposure and leukemia risk

Delzell et al., 1996 conducted a study of 15,649 men who had worked for at least one year (during 1943-1991) at any of eight styrene-butadiene rubber (SBR) plants. The study included all but one small plant of the earlier Johns Hopkins study (a plant that did not begin SBR production until about 1970) and updated the mortality experience at all plants studied, along with conducting a much more detailed exposure reconstruction for these workers (Matanoski, et.al, 1990). The cohorts differed slightly because of somewhat different definitions of eligibility, but, for all practical purposes, the Johns Hopkins cohort is subsumed by the Delzell cohort. Thus, there is essentially only one study of SBR workers, of which the Delzell, et.al, 1996 and Macaluso et al., 1996 reports represent the most recent follow-up-- for up to 49 years with a mean of 25 years.

1		
2	c)	Lumping of lymphohematopoietic tumors
3		The SAB did not feel it was appropriate to "lump" lymphohematopoietic
4		tumors. Leukemia and lymphosarcoma are separate diseases.
5		
6	d)	The Criteria for Causal Inference
7		The Criteria for Causal Inference are biased in their presentation. They
8		should be revised to reflect the above points. It was generally agreed that
9		the SBR process meets the criteria, but the monomer does not.
10		
11	e)	Additional points that should be noted in the report:
12		i) In the total cohort the leukemia rate was somewhat elevated (O/E=
13		48/36.6= 1.31, 95% CI= 1.0-1.7) but the number of excess
14		leukemias was only about 11 (Delzell, 1996). Surprisingly, there
15		was no excess among those hired before 1950 (O/E= 17/16.4=
16		1.04) when one would expect the highest exposures, but there was
17		an excess among those hired during 1950-59 (O/E= 20/10.0= 2.0,
18		CI= 1.2-3.1). Research is ongoing as to whether the risk elevation
19		beginning in 1950 is a function of concomitant exposure to
20		DMDTC (Dimethyldithiocarbamate) in the SBR process beginning
21		in about 1950, but it is premature to judge that hypothesis at this
22		time.
23		
24	ii)	There should be a table to present the actual exposure-response
25		data for the Delzell/Macaluso study of butadiene exposure and
26		leukemia risk, rather than just burying the values in the text (e.g.,
27		Page 7-22, Line 22 and Page 7-25, Line 7). These are among the

1		most important numbers in the report, so they should be
2		prominently displayed.
3		
4	iii)	According to Macaluso et al., 1996, the 0-dose group contained a
5		substantial number of salaried workers but the other groups
6		apparently did not. This could potentially bias the
7		exposure-response risk estimate, and it should be mentioned.
8		
9	iv)	There seems to be a substantial discrepancy between two sets of
10		risk estimates given for this study in the report on Page 7-22 (Line
11		22) and Page 7-25 (Line 7), as shown in the table below. One
12		wonders if the second set (the lower half of the table), which
13		appear to be the one used by the EPA, may be incorrect. Notice
14		that in the upper half of the table, the RR for >0-19 ppm-years is
15		1.1, whereas in the lower half, the two cells that cover the range of
16		>0-19 ppm-years have RRs of 2.0 and 2.1 rather implausible
17		values, especially the RR of 2.0 for <1 ppm-year. The middle
18		range is likewise higher in the lower half of the table (RR=2.4 for
19		20-79, vs. 1.8 for 20-99 ppm-years in the upper half). The RRs in
20		the high exposure range also differed notably: RR=4.5 for 80+
21		ppm-years in the lower half of the table, but in the upper half the
22		RRs for 100-199 and 200+ ppm-years are 2.1 and 3.6. The reason
23		for the discrepancies is not clear; the only difference noted in the derivation
24		of the two sets of estimates is that the upper set adjusted for years
25		since hire and calendar period, whereas the lower one did not. If one
26		of these two sets is not in error, then the discrepant results suggest
27		that the results must be very sensitive to the particular cutpoints,

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confounders used, etc., which would argue for observing caution so as not to extrapolate from the most extreme results. It is of note that most of the analyses reported by Delzell et al., 1996 in their technical report, e.g., Tables 60-62, are similar to the upper half of the table below, rather than the lower half.

Two sets of estimates of leukemia relative risks (RR) reported from the Delzell/Macaluso study in relation to butadiene exposure:

Exposure range					
(ppm-years)	0	>0-19	20 - 99	100 - 199	200+
RR	1.0	1.1	1.8	2.1	3.6
Exposure range				20 - 79	
(ppm-years)	0	<1	1 - 19		80+
RR	1.0	2.0	2.1	2.4	4.5

It is unclear from the EPA report as to how high the correlation between butadiene and styrene exposure levels was in this study. However, one would expect the correlation to be fairly large. (A personal communication from Jeffrey Lewis indicated that the correlation was 0.53.) Since there is also some indication that styrene may be associated with leukemia in this study, partialling out the effect that is attributable to butadiene is problematic, particularly if there is more reliability or accuracy in assigning butadiene exposures than in assigning styrene exposures, or vice versa, since variations in reliability/accuracy could drive the proportion of variance attributed to one chemical vs. the other in regression analyses.

1		There	e is a concern about the effect of peak exposures that was not
2		consi	dered in estimating risk. More is discussed about this issue in
3		Secti	on f below.
4	f)	Speci	ific Comments
5		i)	Page 7-3, Line. 33: The latency period of 10-19 years was left out.
6			
7		ii)	Page 7-5, Line. 14-15: Singling out an intermediate subgroup that
8			gave a suggestive elevation in risk, when subgroups with more
9			exposure did not, is a questionable scientific procedure (i.e., picking
10			and choosing the ad hoc results that support a particular point of
11			view).
12			
13		iii)	Page. 7-20, Line. 1-5 & Page. 7-33, Line. 9-12: It seems curious to
14			report subgroup analyses based on just 3 of the 8 plants, especially
15			when the reason given for choosing them ("three plants who had
16			geometric means of exposure" out of the 7 plants with
17			measurements) seems irrelevant.
18			
19		iv)	Page 7-21, Line 30-32: This says, "When this subcohort was
20			further restricted" Please define the subcohort that is discussed,
21			and also state the endpoint that is being discussed.
22			
23	v)	Page	7-23, Line 25-26: One important feature was that the job-
24		expo	sure matrices (JEM) estimates were specific for calendar time.
25		This	should be noted.
26			
27	vi)	Page	7-31, Line 32-33: It is inappropriate to cite positive findings

- DO NOT CITE OR QUOTE -1 from some earlier follow-up of the cohort when these were not 2 confirmed by the latest follow-up. 3 4 vii) Page 7-33, Line 1-8 and Page 11-5, Line 12-25: In order to 5 present a balanced review of the Matanoski case-control study, the 6 re-analysis by Cole et al., 1993 that yielded a null RR should also be 7 reported, with an indication of how highly sensitive the results were 8 to the particular cutpoints chosen (Acquavella, et al., 1994). The 9 Cole-Acquavella results are mentioned only obliquely, and the 10 discrepancy is not articulated and its implication are not considered. 11 12 viii) Page 7-34, Line 33,35: To be more balanced, the report should 13 indicate that the Meinhardt results were not statistically significant. 14 15 ix) Page 7-36, Line 23: This sentence should be deleted because it 16 suggests that the stop-exposure studies conducted by Melnick et al. 17 (1990) confirm the findings of excess lymphosarcoma among 18 short-term monomer workers in the Divine et al. (1996) study. This 19 statement is incorrect. 20 21 x) Page 7-36, Line 23: There is no way that the stop exposure 22 studied in mice "confirm" the short term worker effect. Long term 23 workers employed at the same time and later did not develop 24 increased lymphosarcomas. 25 3.1.8 Chapter 8 - Pharmacokinetic Modeling 26

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There was consensus that this chapter is out of date. The chapter should be updated and should include the most recently published PBPK models. A major data gap is the lack of modeling of 3,4-epoxy-1,2-butanediol (EBD). A model describing the kinetics of this metabolite would be useful in relating external exposure concentrations to measures of internal dosimetry such as hemoglobin adducts. Such a linkage would facilitate reconstruction of BD exposure profile in exposed humans. A majority of the members were of the opinion that since DEB is very likely involved in BD-induced carcinogenesis, current models which describe this metabolite would be adequate for risk assessment. Other members felt that other metabolites, such as the diol, may play an important role in carcinogenesis at some sites, and that these metabolites needed to be addressed.

The technical quality and comprehensiveness of this chapter could be improved greatly by the inclusion of the most recent relevant literature on butadiene toxicokinetic modeling. Specific suggestions regarding a reorganization of Chapter 8 are offered below. Chapter 8 of the draft document on toxicokinetic modeling does not reflect the current state of knowledge regarding PBPK models for butadiene. Rather, this chapter describes and critiques the initial attempts of several laboratories to independently develop "first-generation" PBPK models for butadiene.

These models were developed often without benefit of critical data such as solubility parameters and metabolic rate parameters. Additionally, definitive data on concentrations of butadiene and its metabolites in tissues of animals exposed to butadiene were largely lacking, precluding rigorous model validation. As such, some of the conclusions that the investigators drew from these early models are no longer relevant. It would be more appropriate to briefly acknowledge these early and important contributions. Then, attention could be focused on a serious critique of the more recent PBPK models (Sweeney et al, 1996, Csanady et al, 1996, Reitz et al, 1996, and Sweeney et al, 1997). These later models represent a significant advance over the previous generation PBPK models in that they describe the kinetics of diepoxybutane, a critical metabolite of 1,3-butadiene.

Inclusion of a discussion of state-of-the-art models for butadiene kinetics would

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significantly strengthen the scientific quality of the document. In particular, the numerous
statements throughout the document indicating that an adequate PBPK model for butadiene risk
assessment is not available should be revised to reflect the current state-of-the-art. At least two
models that describe the kinetics of butadiene, epoxybutene and diepoxybutane were available to
the EPA in 1996 (Csanady et al, 1996, Reitz et al, 1996) and one was published in 1997 (Sweeney
et al, 1997). A majority of the Committee felt that any one of these models, combined with
available in vitro metabolic rate constants could be used to obtain more refined human dose
estimates while other Committee members were concerned about the degree to which some
models were able to describe the available data, the extent to which parameters had to be changed
from measured values to obtain reasonable fits and the accuracy of the low dose extrapolation. As
noted by the Agency, these refined dose estimates could be used in the animal based risk
assessments for both cancer and reproductive toxicology endpoints. In accord with the revised
EPA Cancer Risk Assessment Guidelines and given what is understood regarding the mechanisms
of butadiene toxicity, a more scientifically based risk estimate for environmental exposure to
butadiene is the most appropriate course of action. In chapter 8, concern is expressed that serious
uncertainties exist pertaining to the model structure, parameter values and validation for the
various PBPK models. It is true that each of the models differ somewhat in the details and that the
parameter values chosen by various investigators are similar but not identical. However, these
models are similar enough that any of the models is capable of predicting blood concentrations of
butadiene, epoxybutene, and diepoxybutane following inhalation exposure to butadiene. This
suggests that whatever differences in the underlying model structure exist, these differences are
minor. Most importantly, each of these second generation PBPK models fully describes the
kinetics of diepoxybutane. A majority of the Committee felt that since it is likely that it is
diepoxybutane that is the critical metabolite for initiation of the carcinogenic effects following
exposure to butadiene, inclusion of this metabolite makes these models especially useful for risk
assessment. An alternative view on the Committee was that other metabolites, such as the diol,
may play an important role and should be addressed in developing models predictive of human

1	risk. Also, th	e models of the diepoxide require validation and further development to address
2	parameter an	d structural uncertainty as well as interindividual variability in human metabolism.
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4	The c	conclusions drawn in Chapter 8 need to be revisited in light of information on the
5	kinetics and	PBPK modeling of butadiene that is not presented in the draft report. In the
6	conclusions,	the Agency notes five areas in which more research is needed including:
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8	a)	evaluation of diepoxybutane kinetics,
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10	b)	investigation of the validity of in vitro metabolic data for extrapolating in vivo
11		exposure,
12		
13	c)	clarification of values of various physiological parameters,
14		
15	d)	better characterization of the distribution values for the human metabolic rates, and
16		
17		
18	e)	more measurement of tissue concentrations of metabolites for model validation.
19		
20	Comments of	n each of these areas are noted below:
21	a)	Evaluation of diepoxybutane kinetics
22		Revision of the draft risk assessment to include data published since January 31,
23		1997 will resolve this need. A paper by Valentine et al (1997) reports on the
24		pharmacokinetics of diepoxybutane in rats after IV injection, the most appropriate
25		route of administration for this non-volatile chemical. Sweeney et al, (1997) were
26		able to successfully simulate this data using a PBPK model developed for butadiene
27		epoxybutene, and diepoxybutane. Given the high mutagenic potency of

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diepoxybutane, it is unlikely that kinetic measurements of this chemical will ever be made in humans in vivo. However, parallel studies using lung and liver tissue samples have quantitated the rates of hydrolysis and glutathione conjugation of diepoxybutane in rats, mice and humans (Boogaard, et al. 1996 and Boogaard and Bond 1996) and in vitro studies on the oxidation of epoxybutene have characterized the rate of diepoxybutane formation (Seaton, et al., 1995).

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b) Investigation of the validity of in vitro metabolic data for extrapolating in vivo exposure.

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The validity of the in vitro -in vivo extrapolation for butadiene has been investigated in a manuscript by Sweeney, et al. (1997), in which the in vitro data describing the rates of butadiene oxidation, epoxybutene oxidation, and hydrolysis and glutathione conjugation of epoxybutene and diepoxybutane determined in vitro were used directly in a PBPK model and the model predictions were validated against in vivo data obtained from butadiene exposure of mice and rats. The ability of the model to simulate both epoxybutene and diepoxybutane blood and tissue concentrations following exposure to butadiene points to the usefulness of in vitro parameters in in vivo models. For carcinogenic or potentially carcinogenic chemicals it is unlikely that direct toxicokinetic measurements will ever be made in people. Therefore, the only viable option is to conduct in vitro experiments in which metabolic rates are determined in human and animal tissue samples and then use these metabolic rates in the context of the PBPK model to predict blood and tissue concentrations of both animal species and humans. Acceptance of this in vitro -in vivo extrapolation strategy would serve to encourage the collection of appropriate data with the ultimate goal of incorporating more mechanistic information into the risk assessment process.

- Regarding the choice of model parameters, there is undue concern that all investigators have not used the same values for physiological parameters. All investigators developing butadiene PBPK models have selected specific parameter values from a distribution of widely accepted parameters values for various physiological parameters in these models. Selection of a single value for ventilation, perfusion, blood flow or organ volume is in itself a simplification of the biological system being modeled. Many of these parameters vary for a single individual throughout the day. All of these parameters vary among individuals. The important point is that the selected values are physiologically realistic.
 - d) Better characterization of the distribution values for the human metabolic rates

 This statement comes across as a gratuitous "more research is needed." As noted
 previously, V_{max} and K_m values have been determined for a number of human
 samples. The fact is that the data base for butadiene is particularly robust in terms
 of the number of samples characterized. If the EPA has concerns regarding the
 quantitative and qualitative distribution of human metabolic rates, these concerns
 should be stated much more explicitly and in sufficient detail that specific research
 could be conducted to meet this need.
 - e) More measurement of tissue concentrations of metabolites for model validation

 Over 13 individual inhalation exposures of mice and rats and monkeys to various
 concentrations of butadiene have been conducted. Measurements at steady state
 and post exposure for butadiene and its metabolites have been made in blood and
 tissues. These data have been summarized by Himmelstein et al (1997). The data
 are remarkably consistent across laboratories and consistently point to dramatic

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species differences in metabolism of butadiene. It is really not clear what additional data the Agency might find useful for model validation.

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Specific comments on Chapter 8 follow:

a) Page 8-1, lines 6-7. The PBPK models cited by the EPA in this report should be updated. More mechanistic and sophisticated models are now available including Reitz, et al. (1996), Csanady, et al. (1996), Sweeney, et al. (1997), and Kohn (1997).

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Page 8-2, lines 1-32. The model of Hattis and Wasson (1987) was developed before there was any experimental data on the rates of 1,3-butadiene activation and detoxication in experimental animals and before there was any experimentally determined data for partition coefficients, estimates of solubility that are used in PBPK models. Given the acknowledged limitation of this model, it would be more appropriate to simply mention that this model is one of the first PBPK models to be developed without going into details regarding the model predictions. This is particularly important given the conclusion of Hattis and Wasson that "differences in pharmacokinetics failed to account for differences in carcinogenesis between mice and rats and that with respect to risk assessment, uncertainties in PBPK modeling are trivial compared with the differences in apparent sensitivities between these species" (Page 8-2, lines 18-32). The overwhelming body of experimental data on the toxicokinetics of 1,3-butadiene and its metabolites collected in rats and mice since the publication of the Hattis and Wasson model have shown that differences in pharmacokinetics between these two species can account for species differences in carcinogenesis. It is also significant that the Hattis and Wasson model was an unpublished report and never benefited from peer-review. It would be more appropriate for the EPA to devote a paragraph to the presentation of the Hattis and

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1 Wasson model similar to what was done for the Hallenbeck (1992) model (page 8-4, 2 lines 3-9). 3 4 Page 8-4, lines 10-33. It might be appropriate for the EPA to cite the Kohn c) 5 (1997) model instead of the 1993 model primarily because the model of Kohn and 6 Melnick (1993) relied on theoretically derived partition coefficients rather than 7 experimentally determined partition coefficients. The use of these calculated 8 partition coefficients resulted in overpredictions of the concentrations of 9 1,3-butadiene in tissues, especially fat. 10 11 d) Page 8-7, lines 27-34. As noted above, the use of empirically derived calculated 12 partition coefficients rather than experimentally determined values led Kohn and 13 Melnick in their 1993 model to conclude that storage in fat is a significant fraction 14 of the retained 1,3-butadiene, especially in rats and humans. It is generally 15 recognized that this conclusion is based on the use of calculated rather than 16 experimentally measured partition coefficients. Inclusion of this paragraph in the 17 chapter may lead readers not knowledgeable in 1,3-butadiene toxicokinetics to 18 believe that this model prediction is accurate. While the EPA does discuss this 19 limitation (page 8-8, lines 1-13), it is still misleading for the EPA to devote 20 significant discussion to models in which theoretical values are used when later 21 experiments determined the values to be inaccurate. It would be much more 22 prudent for the EPA to report the most recent PBPK models noting when 23 necessary that these laboratories had also participated in the development of earlier 24 models. 25 26 Page 8-12, lines 21-23. The EPA notes that "Johanson and Filser are reportedly e) 27 working on a corresponding PBPK model for humans but it has not yet been

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published." At this point it would be most appropriate for the EPA to cite the Csanady et al (1996) PBPK model for 1,3-butadiene that includes model predictions for man.

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f) Page 8-15, lines 9-11. The EPA notes that a limitation of the model of Evelo et al., 1993 is that metabolism of butadiene is limited to the lung and the liver. This should not be viewed as a limitation. The objective of PBPK models is to account for the most significant mechanistic steps in the disposition of chemicals with the goal of predicting the concentration time profile of the toxic agent in either the target tissue or a suitable surrogate for the target tissue such as blood. It is not practical nor is it necessarily advantageous to develop a model that incorporates all pathways in the disposition of a chemical however minor. The simplest models that are the most useful for risk assessment are most likely to have the greatest value since these models will have the fewest number of parameters that require independent experimental determination in animals and humans. Thus, it is not clear why the Agency feels it is necessary in the case of butadiene to account for metabolism of this chemical in all tissues of the body. Extensive modeling efforts have determined that inclusion of metabolic activation in the liver, the major organ for metabolism of butadiene, and in the lung, a target organ for mice, are the most appropriate from a mechanistic standpoint.

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g) Page 8-15, line 15. The EPA notes that the most recent PBPK model published for 1,3-butadiene is the model of Medinsky et al. (1994). This statement is correct only in the context of the January 31, 1997 cutoff date for consideration of reports in this document. However, the statement is misleading as there are a number of other PBPK models that have been published since 1994, including models published in

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1996 and 1997. The EPA is urged to revise this chapter to include these more recent models. These models not only consider the metabolism of 1,3-butadiene, but also include the disposition of its two epoxide metabolites, epoxybutene and diepoxybutane, thereby making the models more appropriate for use in risk assessment.

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Page 8-16, lines 6-14. The EPA notes that "in the model of Medinsky et al., the microsomal concentrations reported by Csanady et al. were not used to scale metabolic rates. Instead, literature values for microsomal concentrations were used." While the Agency correctly summarizes the approach taken, the rationale for taking this approach is not appropriately presented. The objective of the study reported by Csanady et al. (1992) was to determine the rates of butadiene oxidation in microsomes from rodents and humans. To achieve this objective it is essential that purified microsomes be obtained from liver, but it is not essential that all of the microsomal protein in the liver be accounted for. Thus, Csanady et al., 1992 report microsomal yield which is less than the total microsomal protein content of liver or lung. Other investigators, in contrast, have sought to determine the total amount of microsomal protein in liver or lung. These investigators have used other techniques for this assessment. Extrapolating in vitro results expressed per milligram of /microsomal protein to the entire organ requires knowledge of the total amount of microsomal protein in an organ not simply the yield of microsomal protein obtained in the biochemical experiment. Thus, the appropriate approach for scaling in vitro rates to the whole animal is to use total microsomal total protein content rather than yield. The literature values used by Medinsky et al. (1994) for total microsomal protein content were similar to those used by Johanson and Filser (1993) and Kohn (1997). In contrast, Kohn and Melnick (1993) used the values of microsomal yield reported by Csanady et al. (1992).

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i)	Page 8-19, Section 8.3, Summary. The EPA notes that "pharmacokinetic
	modeling of 1,3-butadiene has not elucidated the reasons for interspecies
	differences in carcinogenic response between rats and mice." This statement is not
	true if one evaluates the current PBPK models that describe the disposition of
	1,3-butadiene, epoxybutene and diepoxybutane. These PBPK models clearly
	demonstrate that the dramatic species differences between rats and mice in
	response to 1,3-butadiene are most likely related to species differences in rates of
	formation and removal of the diepoxybutane metabolite. Experimental data and
	PBPK model simulations indicate that mice produce far greater concentrations of
	this reactive metabolite compared with rats. Sweeney et al. (1997) have used in
	vitro metabolism data collected in tissues from rats, mice, and humans directly into
	a PBPK model to make predictions regarding the epoxide concentrations in blood
	and tissues following exposure to 1,3-butadiene. Using in vitro derived parameters
	they were able to adequately simulate the pharmacokinetics of 1,3-butadiene,
	epoxybutene, and diepoxybutane. This ability to use in vitro data to make in vivo
	predictions suggests that in vitro data on rates of 1,3-butadiene, epoxybutene, and
	diepoxybutane metabolism obtained in human tissue samples can also be used to
	predict blood and tissue concentrations of both epoxide metabolites in humans.
	When these simulations are conducted, using average values for human metabolic
	rates, the results indicate that diepoxybutane concentrations in humans would be
	orders of magnitude less than those of mice and lower than, but much more similar
	to, concentrations predicted for rats. Thus, the EPA should revise this chapter to
	include a discussion of the toxicokinetic models for 1,3-butadiene that are now
	capable of simulating not only the disposition of 1,3-butadiene, but also its two
	most important epoxide metabolites.

j) Page 8-19, lines 33-34. The EPA statement that uncertainties in the existing

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PBPK models and data make them unreliable for use in risk assessment must be revisited. Given that several investigators have been able to use these models and the underlying metabolic data to predict butadiene, epoxybutene, and diepoxybutane concentrations in rodents exposed to butadiene, it is not clear what additional data and what uncertainties need to be resolved prior to the use of these models in risk assessment. For example, Sweeney et al. (1997) have used in vitro metabolism data obtained from rat, mouse, and human tissues directly in a PBPK model to make predictions regarding the epoxide concentrations in blood and tissues following exposure to 1,3-butadiene. Using in vitro derived parameters—they were able to adequately simulate the toxicokinetics of butadiene, epoxybutene, and diepoxybutane.

k) Page 8-20, line 5-21. In this paragraph the EPA presents a number of criticisms regarding the parameters used in the PBPK models that are not necessarily accurate. For example, the EPA notes that "with respect to parameter values, there are disagreements about the ventilation rate and about metabolic parameters." As noted above, while it is true that each investigator uses a different value for ventilation rate or metabolic parameter, the values used by all investigators are within the normal range associated with these parameters. It is also not unreasonable to expect that if the EPA were to use a PBPK model in risk assessment that other point estimates also within the range of reported values for these parameters would be chosen by the EPA.

The EPA also notes that there is a paucity of human in vitro data for extension of the PPBK model to humans and that the few measurements that have been made on a few metabolic parameters show a high amount of variability. Relative to other chemicals there is an extensive amount of experimental data on the rates of

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metabolism of 1,3-butadiene by human tissue samples. Each of the metabolic pathways 2 important in the disposition of 1,3-butadiene and its metabolites have been 3 quantitated with V_{max} and K_m values obtained from multiple human samples. These pathways include oxidation of 1,3-butadiene, oxidation of epoxybutane, hydrolysis of epoxybutane, glutathione conjugation of epoxybutane, glutathione conjugation of 6 diepoxybutane, and hydrolysis of diepoxybutane. Means and standard deviations for these parameters have been calculated because multiple human samples have been used. Thus, sample distributions can be generated from which population values can 8 9 be obtained. A PBPK model applied to human risk assessment could employ either 10 average values to obtain deterministic predictions or Monte Carlo simulation techniques to get probabilistic estimates of the range of responses of hundreds of 12 simulated humans. The latter approach using Monte Carlo simulation would 13 provide some estimate of the potential variability in human response to inhaled 14 1,3-butadiene in addition to an estimate of the response of the most sensitive humans. 15 Regarding the large amount of variability associated with these metabolic parameters when measured in humans, this reflects the inherent variability in the 16 17 expression of various xenobiotic metabolizing enzymes in the human population, a 18 fact that has been well documented in the literature. Thus, it is not unexpected that 19 of metabolic capacity from tissues from multiple humans should assessment 20 yield a range of outcomes. 22 1) Page 8-20, lines 22-34. The EPA notes that the existing models have been

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subjected to a very limited validation mostly by comparison of simulation results with chamber uptake data. This statement is true regarding the PBPK models that were published in 1994 and earlier (e.g. the models reviewed in this current document). However, since the publication of these first generation models, multiple inhalation toxicokenetic studies have been conducted in rats and mice

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where blood and tissue concentrations of 1,3-butadiene, epoxybutene, and diepoxybutane have been quantitated following inhalation exposure to 1,3-butadiene. The second generation models that include the prediction of not only epoxybutene but also diepoxybutane have utilized this recent in vivo toxicokenetic data for model validation.

The EPA also notes that "for PBPK models to be more reliable, they should also be validated against tissue concentration data for various metabolites and various tissues. More recently these data have become available although they must be interpreted with caution because it appears that metabolites in some of the tissues are subject to further metabolism during the lag time between termination of exposure and measurement of tissue concentrations." This statement is true and ironically the implications of post exposure metabolism were first recognized when one of these second generation PBPK models (Sweeney et al., 1996) failed to adequately simulate tissue concentrations of epoxide metabolites. When the authors modified the initial conditions of the model to reflect the time lag between termination of exposure and measurement of tissue concentrations and the capacity of the tissues to metabolize 1,3-butadiene post exposure, they were successful in predicting the actual measured epoxide tissue concentrations.

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Page 8-21, lines 4-23. The EPA concludes that the existing PBPK models and data cannot explain the interspecies differences in 1,3-butadiene carcinogenicity. As noted previously, the first PBPK models discussed in this report did not include the toxicokinetics of diepoxybutane. More recent second generation PBPK models that include the formation and elimination of this metabolite are successful in simulating in vivo data in both rats and mice for 1,3-butadiene for epoxybutene and diepoxybutane concentrations in blood and tissues. Both model predictions and experimental data indicate that the dramatic interspecies differences in carcinogenic

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1		response to 1,3-butadiene can, in fact, be explained by the dramatic interspecies			
2		differences in circulating concentrations of the diepoxybutane.			
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4	3.1.9	Chapter 9 - Quantitative Risk Assessment for 1,3-Butadiene			
5					
6	The A	agency is to be commended for its efforts to develop a clear, well-reasoned			
7	quantitative a	ssessment of 1,3-butadiene cancer risk. The assessment is made particularly			
8	difficult, by the	ne continuing release of new and relevant scientific data, including toxicokinetic,			
9	epidemiologi	cal and mechanistic data. Furthermore the analyses were undertaken after the release			
10	and SAB review of the proposed EPA carcinogen guidelines but before they have been finalized.				
11	The Agency is to be commended for looking at new approaches, such as the benchmark dose				
12	procedures, to improve quantitative assessment on non-cancer endpoints. However, the				
13	Committee has submitted suggestions on how to further improve these approaches and how to				
14	make these new approaches more clear, accurate and consistent. Specific comments on chapter 9				
15	are given belo	OW.			
16					
17	a)	Dose response analysis of the human carcinogenesis data			
18					
19		i) Quality of human data			
20					
21		The Committee agrees that the Agency's risk assessment of 1,3- butadiene			
22		should include the latest follow-up of each of the epidemiologic studies, including			
23		Delzell et al. (1995) and Matanoski (1997). Delzell fits a variety of dose response			
24		models, accommodating a wide range of dose response curves. The models fit equally			
25		well, and as noted by EPA as a whole were consistent with a considerable range of low			
26		dose risk predictions. Thus the Delzell study does not permit one to discriminate			
27		among models for the purpose of low dose prediction. In this sense the data can be			

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viewed as limited for the purpose of low dose risk prediction. The reason why equivalent model fits are obtained requires further explanation in order to convey a greater appreciation of the nature of the dose response data. For example, is this due to a relatively narrow range of dose response data in the exposed cohort or to considerable scatter?

The assessment notes that few subjects were exposed to benzene, but benzene was not seen to confound the relationship between butadiene or styrene and leukemia. Because benzene is widely recognized as a human leukemiogen, further discussion of this point is needed (e.g., an indication of whether the benzene exposures were insufficient to cause effects; consistency with expectations based on a Chinese cohort).

In the section on uncertainties, the potential importance of exposure misclassification in the Delzell study is discussed, but further discussion of the uncertainties of dose reconstruction and the potential magnitude of impact on the risk predictions is desirable.

ii) Presentation of multiple model fits and low dose extrapolations

The results of the multiple model fits derived by Delzell and colleagues are tabulated, along with low dose risk predictions. While it is reasonable to compare results to make the point that the data do not provide the basis for discriminating among models of quite different shapes, once it is established that this is the case the further presentation of low dose analyses is not needed and unnecessarily complicates the presentation (e.g., in Table 9-3 and Table 9-4). Presentation of results for the linear model would suffice, along with perhaps effect concentration estimates, within but

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not outside the range of observation. The other point to be made, that the exposure concentration at 1% risk level (EC₀₁₎ results for the final versus initial square root model differ by roughly 40 - 50% could also be made, for example, in the text. It motivates the need for obtaining comparable estimates for the linear model, by omitting styrene and race (as was done for the "final" square root model).

Included are the results for the square root model, as well as the power model, which is apparently best fit by a coefficient less than unity. Superlinear models such as these are not biologically plausible, and should therefore not be used for low dose risk predictions. Synergy between genotoxicity and cell proliferation frequently occurs as does saturation of detoxification and metabolic activation. These factors result in sublinear data sets. Metabolic data for butadiene clearly show saturation of activation and the molecular dosimetry data are supralinear. An analogous point was made by the Committee in commenting on proposed EPA carcinogen guidelines regarding the use of the Weibull-in-dose model without constraints for exponents less than unity for risk prediction. Lack of biological plausibility is another reason for not presenting low dose risk predictions for the square root and power models.

The measure of deviance for the models seem to be within 0.4, while normally one would say models differ if their difference in deviance is about 4.0. Also, the best fit for the multiplicative model is sublinear; for the power model it is superlinear and as noted by Delzell and colleagues, the square root model fits best (although marginally so) - again, this reinforces the notion that the data provide limited information on dose response. Perhaps replacing the dose response figures with one figure showing the data points with their confidence intervals, and superimposing on the figure the various models fit would provide a better

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understanding of why this is the case. The inclusion of data should clearly illustrate why the data are equally explained by the wide range of models tried.

iii) Dose rate, duration and timing

With benzene induced leukemia and lymphoma the recency of exposure is an important feature of the dose -time-response, with recent exposures most important for leukemia, and distant, high exposure most important for lymphosarcoma. On Page 9-2, the assessment notes the finding of Delzell and colleagues that excluding exposures within 5-10 years of death slightly increased the exposure response relationship but excluding exposures within the last 20 years almost eliminated the relationship. It would be desirable for the Agency to explore in detail the possible dose time relationships. On a related point, Delzell and colleagues have recently reported on the possible importance of peak exposures to 1,3-butadiene on leukemia risk (Delzell, et al., 1995). It is important to explore the dose rate/time/response issues through careful analyses of the Delzell data set.

iv) Extrapolation of occupational results to general population

The available data for risk assessment is for males, exposed as young and middle-aged adults, and for a selected group in that they were workers in the jobs studied. The extent to which this group can be used to represent the general population should be addressed more carefully, with an attempt to quantitatively address, where possible, differences between cancer potency for the occupationally exposed and the general population. Such an analysis should consider the following:

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1	a)	potential increased sensitivity for women and girls
2		No dose response information appears to be available for women.
3		The mammary gland was a sensitive site in both rats and mice.
4		Given the large public health concern for breast cancer in women,
5		and the findings in the bioassay, the issue should be explicitly
6		addressed. In terms of overall risk, female rats in the Hazelton 1981
7		study were clearly more sensitive than male rat, at least by an order
8		of magnitude. Although given the uncertainties in the quantitative
9		analysis, this is less clear for the mouse, cancer potency estimates
10		were greater in the female compared to the male by about a factor
11		of 2. Although toxicokinetic differences are not apparent in
12		hemoglobin data for men and women with similar exposure, the
13		potential for increased susceptibility of mammary tissue cannot be
14		excluded.
15		
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17	b)	healthy worker effect
18		The extent to which a presumably small adjustment may be appropriate
19		should addressed. The extent to which restrictions on smoking in
20		occupational settings with butadiene exposure may have contributed
21		to the healthy worker effect should be addressed by the Agency.
22		There was a diversity of opinion amongst the Committee on this issue.
23		Internal comparisons done by Delzell et al. (1995) that were
24		used to derive the human cancer potency estimate are not influenced
25		by the healthy worker effect since no external comparison population
26		is being used. However, the healthy worker effect is an important

consideration in extrapolating from the occupational setting to the

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1		general population, and this could not be assessed through an internal
2		comparison.
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4	c)	exposure during a working life versus other life stages
5		Issues falling under this category to address include the potential
6		for inherent increased susceptibilities at different ages (e.g., in
7		utero, and during infancy, childhood and old age), and those related
8		to the stages and mechanisms of carcinogenesis (e.g., time of
9		exposure versus observation; after a point, diminution of risk with
10		time since exposure). Analyses of the Delzell and perhaps
11		Matanowski et al. 1997 data, and observations from the large
12		Chinese benzene/leukemia cohort may shed light on the second
13		issue. The available data on butadiene obviously does not provide
14		direct information on the first issue, but data on leukemogenesis
15		from other agents may.
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18	d)	other potentially susceptible subpopulations
19		The potential range of susceptibility within the general population
20		should be explored quantitatively, by for example, considering the
21		impact of polymorphisms on low dose risk, and addressing the extent
22		to which certain potentially susceptible groups are contained within the
23		Delzell cohort.
24		
25	The document would	be improved by presenting greater detail on the derivation of lifetime
26	risks from continuous	exposure from the fits to the occupational data. Perhaps this could
27	be done in an appendi	X.

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v) Choice of maximum likelihood estimate over Upper Confidence Limit (UCL) on potency/LEC(95% lower confidence interval)

Maximum likelihood estimates (MLEs) rather than lower confidence bound

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on the effective concentration (EC) (or upper confidence bound on potency) are used in the final characterization of the dose response derived from the human data. It has been the practice of the Agency to use maximum likelihood estimates in potency derivations from human data and upper confidence bounds on derivations from animal data. The rationale provided in the 1,3-butadiene document is that the simple linear model fit to the human data does not have the instabilities that can be associated with polynomial fits, and that there is far less uncertainty in the potency estimate derived from human data. In this particular case there are a number of unaddressed issues suggesting that, at least, the EC₀₁ may be underestimated for the general population. These include exposure misclassification in the Delzell cohort, the lack of availability of data on all but males exposed occupationally during adulthood, and the lack of availability of a fit of a "final" linear model (comparable to the model for the square root model [omitting styrene and race]). In addition, as previously stated in this report, there is a possibility that there was an overestimation due to exposures being underestimated in the Delzell et al. (1995) study. It would be preferable to explicitly take these factors into account, to the extent possible; it is unlikely that application of a confidence bound will provide adequate correction.

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vi) Availability of Delzell study

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Although the Delzell study has been published (Delzell, et al., 1996; Macaluso, et al., 1996), some of the key analyses that aid in interpreting the data

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1 are available only in an unpublished technical report and thus is of limited availability 2 to the public. At the meeting the Committee heard that there was peer review of the 3 unpublished technical report. A report so critical to the dose response analysis and 4 so heavily cited in the document should be made widely available to the public, along 5 with description of the peer review conducted by the Agency. 6 7 8 b) Dose response analysis of animal cancer bioassay data 9 10 i)Time dependent analyses 11 12 The default, time dependent analyses of the cancer bioassay data were 13 carefully done and are well presented. 14 15 Because the cancer incidence rates are large for some tumor sites, errors 16 are introduced if it is assumed the polynomial represents a true model of multistage 17 carcinogenesis, as Suresh Moolgavkar (1994) has pointed out (Moolgavkar, 18 1994). This should be acknowledged in discussions which infer number of stages 19 on the basis of modeling results (e.g., Page 9-20). The errors may be relatively 20 small given the availability of data at relatively low doses; it would be desirable to 21 attempt to gain an understanding of the magnitude of the error for these fits. 22 23 The method used to develop the upper 95% confidence bound for the sum 24 of the incremental lifetime unit cancer risk for humans (q_1) parameter across sites is 25 reasonable. An alternative approach for future assessments would be to derive the distribution for the q₁ parameter for each tumor site and use Monte Carlo 26

simulation or numerical techniques to obtain the distribution of the sum. The result

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should differ marginally from the one presented, and thus it is not been suggested that the analysis be redone for this case.

ii) Lack of site concordance across species

While leukemia is the observed endpoint in humans, leukemias are not observed in the animal bioassay in either rats or mice, in either sex. The rationale for the default inferences regarding site concordance, and the inclusion of sites such as the Harderian gland and forestomach should be clearly stated. Ovarian tumors are observed, along with ovarian atrophy in the animal bioassay. The degree to which the toxicity may have played a role in certain sites observed in the bioassay and the extent of relevance to humans exposed at lower doses should be discussed.

iii) Toxicokinetics

A variety of viewpoints were expressed within the Committee over the extent to which toxicokinetic analyses should be incorporated into the assessment for use in interspecies and high to low dose extrapolation. The Agency was criticized by some for not incorporating the results of the recent models. Others noted significant deficiencies in the proposed models and the very recent developments suggesting the field was undergoing rapid development, and found the models speculative. Nonetheless, the different toxicokinetic hypotheses and the hypothesized role of the various metabolites should be discussed, and at least qualitatively, with an indication of the degree to which the assessment would be impacted if some of the hypotheses were later proven true. The discussion should acknowledge the variety of competing viewpoints and hypotheses of

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1	re	searchers in the field. Where information on toxicokinetics and mechanism provide					
2	ad	equate understanding of findings at certain sites within one rodent and not the other,					
3	th	is should be given.					
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6	c) Th	ne assessment of the developmental and reproductive toxicity endpoints					
7							
8	In	approvement in non cancer risk assessment, such as use of effect levels as suggested in the					
9	benchmar	k dose procedures, was supported by the Committee. However, since these are					
10	relatively	new procedures, the Agency must meet a high standard of clarity and transparency in					
11	their initia	their initial applications as presented in this 1,3-butadiene assessment. The Committee encourages					
12	the EPA t	the EPA to review the accuracy of their calculations. The Committee also requests discussion in					
13	the risk assessment EPA to explain the use of their newly proposed models, especially those						
14	modeling time to impact and to provide additional explanation of the safety factors applied to the						
15	benchmar	k calculations resulting from their modeling exercises:					
16							
17	i)	A clear rationale for the selection of one particular point of departure over another					
18		is needed for each case for which the default is applied and for cases deviating					
19		from the default.					
20							
21	ii)	Where it is not obvious the biological significance of particular endpoints for the					
22		assessment should be conveyed.					
23							
24	iii)	When applying novel approaches, such as the time-to-response modeling of					
25		atrophy, the advantages and limitations in utilizing the approach in the context of					
26		reference concentration estimation should be clearly understood and presented.					
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1 iv) The incorporation of information on toxicokinetics (e.g., ovotoxicity of diepoxide) 2 should be used where possible on at least a qualitative if not quantitative basis to 3 inform the choice of endpoints, modeling and uncertainty factors. 4 v) The exclusion of points at high doses contributing to a poor fit is a reasonable 5 approach, and typically can be explained, for example, in terms of toxicities other than 6 the one being modeled, or toxicokinetics. When the approach is applied where 7 possible the specific rationale for the case at hand should be presented. The use of the 8 procedure and the rationale provided for should be consistent across endpoints and 9 across assessments for the same scenarios. 10 11 There may be a difference of opinion among the EHC about the appropriateness of vi) 12 using a risk reduction factor, when reference concentrations are based on 13 benchmarks associated with risk of effect. The rationale for use of this factor needs 14 to be clearly laid out. 15 16 vii) Analyses of this type applied to these endpoints may be unfamiliar to others 17 working in the general area. Confusion over the presentation of some results 18 (e.g., logarithm versus linear scales) was obvious in some of the comments 19 received. Also, calculations should be easily followed. Some of the Committee 20 noted, as did several consultants presenting comments to the Committee, that 21 mathematical errors were evident. EPA should carefully review this section of the 22 document. Tables and figures should be free standing, with statistics referenced. 23 24 Additional comments recommendations for Chapter 9 include the following: Page 9-4: Since previous EPA lifetime risk estimates have used a 70-year 25 a) time-frame, it would seem appropriate to follow this precedent for the sake of 26 27 comparability, rather than use an 85-year estimate.

DRAFT #4 - DRAFT DOCUMENT-- DO NOT CITE OR QUOTE -1 Figures 9-9 to 9-14: These figures should be prepared so that the axis are b) 2 understandable without reading the full text. 3 c) Page 9-27, line 33: Why does this sentence say 600 ppm and above? Why doesn't it say 625 ppm, a dose that was tested? 4 5 6 d) Page 9-5 & 9-7: There is a discrepancy in the 95% lower confidence interval with 7 a 1% level of risk, LEC₁, value between the text and the figure (0.12 ppm vs. 8 0.066 ppm). Two of the figures appear to be mislabeled. 9 10 Page 9-13, lines 18-19: It was difficult to follow the logic in the last sentence. e) 11 This sentence should be modified to provide necessary rationale to follow this 12 choice. 13 14 15 f) Page 9-36, lines 13,14: What is the rationale for this statement? Either the explanation should be expanded or the statement should be deleted. 16 17 18 Tables 9-13 thru 9-15: Statistics should be included for these summary tables. g) 19 20 Figures 9-9 thru 9-14: For clarity, the axis should be labeled so that reader can h) 21 easily convert dose to ppm without going back to the text. Also figures, like tables, 22 should indicate when exposures were adjusted to 24 hour daily exposures. 23 24 i) Section 9.3: Although the 95% lower confidence interval with a 10% level of risk

(LEC₁₀) values are taken for the "point of departure" for the RfC's calculated for

was chosen over the 95% lower confidence interval with a 5% level of risk (LEC_s)

LEC₁₀

the benchmark based approach, minimal discussion was given as to why the

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1		nor why the 95% lower confidence interval with a x% level of risk (LEC _x) versus ED
2		values are chosen. Justification and rationale for this issue for all modeled endpoints
3		should be provided.
4	j)	Pages 9-49, lines 9-13 and Table 9-21: The Committee was unable to follow the
5		logic in the use of the "risk reduction factor" that was applied to the benchmark
6		dose based RfC. This section should be rewritten so that it is comprehensible.
7		
8	k)	Pages 9-49, lines 33-34: Is this statement true for dominant lethal effects as well
9		as fetal weight reduction? This Committee urges caution in such general
10		statements or authors should provide specific justification of these statements.
11		
12	1)	Pages 9-42, lines 5-7: Text cites Allen et al. 1994b as source of information
13		supporting the use of LEC ₁₀ as being "at or below the range of detectable
14		responses." (Allen, 1994) Since this paper dealt only with developmental toxicity
15		data, this statement as well as those later in this section are extrapolations from
16		that research. The following text should be reworded, "Other studies are
17		supportive of this statement. For example, the statistical power of detection of this
18		study design supports this statement."
19		
20	m)	Pages 9-44, Table 9-19: A footnote should be added to describe what the Z
21		statistic is and how Q O-2 are obtained. This section was very unclear.
22		
23	n)	Pages 9-40 thru 9-44, Section 9.3.4, pp. 5-3 thru 5.5 and Section 5.1.4: The
24		Agency needs to address more fully the statistical and biological significance of the
25		testicular atrophy. The footnote on page 5-4, Table 5-1 states that statistics were
26		not conducted on the testicular lesions yet in Section 9.3.4 this endpoint becomes a
27		study for modeling. Is the background rate for this lesion in the control B6C3 F1

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1		mice low compared to historical mouse population statistics?
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3	o)	Pages 9-46, line 1-2: In this example, the text provides a reason for discarding the
4		top doses in the modeling however, this decision is inconsistently applied in the
5		modeling Section 9.3 as evidenced in Table 9-16. The text should provide some
6		common guidance on what will be done regarding dropping higher dose levels
7		from modeling calculations.
8		
9	p)	Pages 9-46, line 10-11: Statement in lines 10-11 appear to differ from more
10		recent research published on PBPK models. Can this health risk assessment go
11		further in using the data about known ovotoxicity of the diepoxide? See examples
12		provided in testimony from the Chemical Manufacturers Associations's documents.
13		(CMA, 1998 a,b)
14		
15	q)	Pages 9-51, lines 2-4: The Committee was unconvinced about the superior nature
16		of the time-to-response modeling that was conducted for the LEC ₁₀ determination
17		for atrophy. Additional discussion is needed to support this statement especially
18		given the limitations of the biological time-to-response data for this endpoint.
19		
20	r)	Pages 9-51, lines 16-26: The EHC agrees with these limitations but would then
21		use these points to justify using PBPK modeling to improve the target organ and
22		time concentration curves relative to these specific reproductive versus
23		developmental endpoints. The text has provided the justification but the
24		assessment falls short of acting on these suggestions.
25		
26	s)	Pages 9-51, lines 27-31: The Committee agrees on these issues. Please see the
27		earlier comments on how to constructively address these points.

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t) Mathematical errors need to be corrected.

It is absolutely necessary that all calculations in Section 9 be carefully reviewed. The Committee noted numerous problems in this section and refer USEPA to the written testimony of R. Seilken that is especially relevant to this issue (Seilken, R., 1998). There may be confusion with some numbers presented as natural logs versus non-log numbers. Please proof very carefully. All calculations must be easy to follow by a general scientific audience. Chapter 9 did not meet this goal.

3.1.10 Chapter 10 - Weight of Evidence

Cancer endpoint

There was a range of opinion on the committee about whether one can conclude that 1,3-butadiene is a known human carcinogen. There was consensus on the Committee that there is sufficient evidence to say that working in synthetic rubber production is causally associated with leukemia. However, the majority felt that there were conflicting results among SBR and monomer workers for leukemia and a lack of compelling evidence for a relationship between lymphosarcoma and butadiene exposure. There was concern that an increased risk of leukemia was not seen in the epidemiology studies of workers in the butadiene monomer industry. To some EHC members, this lack of association significantly reduced confidence in the assumption that 1,3-butadiene was the causative agent for leukemia in the SBR industry study. In addition, confounding or coexposure to other chemicals could not be ruled out with confidence.

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There were a variety of views amongst the EHC as to how the call should be made on whether a substance can be considered a "known human carcinogen." One view was that the human data - from observational studies of cancer in humans - must stand on its own to make the finding of "known human carcinogen", without regard to mechanistic or other information. A second view was that the cumulative evidence, from human and animal studies, as well as mechanistic data, particularly as it relates to human findings, should be used as the basis for the judgment. The majority of the Committee felt that the judgment should be made on the basis of human cancer observations alone and the evidence was not sufficient for 1,3-butadiene. A few of the Committee members considered the human evidence in and of itself sufficient or that the cumulative evidence was sufficient to make a finding of "known human carcinogen." Other Committee members considered the body of mechanistic data to be indicative of the fact that significant interspecies differences in response to 1,3-butadiene exist between rodents and humans. For these members, the mechanistic data were consistent with 1,3-butadiene not being classified as a known human carcinogen.

With respect to the narrative discussion in the evaluation, the Committee felt that it should more reflect the range of opinion on the matter when discussing the human findings. Clearly 1,3-butadiene, when mutagenic or clastogenic, is so through its metabolism. This finding should be provided in the evaluation, as well as a statement regarding the mechanistic studies most relevant to humans.

Non-cancer endpoints

The weight of evidence is confined to addressing cancer endpoints. The reproductive endpoint is the basis for RfC calculations and, therefore, also should be addressed.

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3.1.11 Chapter 11 - Risk Characterization

The majority of the Environmental Health Committee did not classify 1,3-butadiene as a known human carcinogen due to the lack of consistency between exposure and leukemia or lymphosarcoma data when the styrene-butadiene rubber (SBR) and monomer worker studies were considered in total. The majority opined that 1,3-butadiene should be classified as a probable human carcinogen where as a minority felt that the science supported the classification of 1,3-butadiene as a known human carcinogen. The vast majority of the panel felt that the SBR process supported classification as a known carcinogen.

The majority of the EHC was reluctant to classify 1,3-butadiene as a known human carcinogen because it was felt that there was no consistent relationship between exposure and leukemia or lymphosarcoma when the SBR and monomer studies were considered in total. Only one study population (SBR industry) had credible leukemia excess related to exposure. Leukemia was not elevated or related to estimated exposure in butadiene monomer industry. Lymphosarcoma was only elevated in short term, not long term workers in the monomer industry and also not elevated in the SBR industry. Hence, there is no dose response for the lymphosarcoma. The lymphopoietic cancers should be considered separately when assessing consistency across studies. The majority of the Committee felt that the finding of "known human carcinogen" should solely be based on observational studies in humans, without regard to mechanistic or other information. Others on the Committee felt that 1,3-butadiene should be identified as a "known human carcinogen" because the cumulative evidence, from epidemiology, animal cancer bioassays, and mechanistic studies should be used as the basis for the judgment.

In substantiating EPA's classification of 1,3-butadiene as a carcinogen, it is important that the following points be made so that the strengths and weaknesses of the data are discussed.

a) Page 11-1: The statement regarding the lack of sufficient data to determine if

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1		children or other subpopulations are affected differently by exposure to
2		1,3-butadiene should be revisited in light of the studies by Nelson et al., 1995,
3		Wiencke, et al, 1995. Nelson et al., 1995 found the glutathione transferase theta
4		(GSTT1) to be highly polymorphic wide variation in its ethnic distribution (Nelson
5		et al., 1995). Wiencke et al., 1995 have shown the association between the
6		genotype for the production of glutathione transferase theta and genetic anomalies
7		in sister-chromatidic exchange induced by metabolites of 1,3- butadiene:
8		epoxybutene and diepoxybutane. This implies that the protein produced by the
9		gene is important in conjugating both of these metabolites. GSTT1 only affects
10		DEB and GSTM1 only affects EB. These are two proteins from two genes. Given
11		the profound racial distribution of the polymorphism, it is important to note that
12		this may account again for the significant portion of the alterations in the
13		metabolism of 1,3-butadiene. Consequently, the enzymes responsibilities for the
14		metabolic conversion of 1,3-butadiene to its mono- and di- epoxide forms as well
15		as its diol form are highly polymorphic. This implies that there may be differential
16		susceptibility to the genotoxic effects of exposure to butadiene. Investigations of
17		this are rapidly moving ahead. There is some indication from field studies that
18		these polymorphisms may contribute to directly- measurable genetic effects (Sorsa
19		et al). Hence, risk assessment and future studies of this compound should take note
20		of this and adjust as is appropriate.
21	b)	Page 11-3, line 11-13 and Table 11-1: EPA states that the conclusion of
22		"sufficient evidence" of human carcinogenicity is based on more than 10
23		epidemiologic studies examining five different groups of workers. This statement is
24		misleading because it implies that there is a consistency of results across several
25		studies of equal caliber examining completely different populations. The
26		predominant emphasis should be on the methods and findings of the two latest

studies of butadiene monomer and SBR workers. The emphasis on the early

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epidemiologic studies is misleading to the extent that it gives the impression of 2 consistency across several study populations in the SBR and butadiene monomer 3 industries. There is really only one study population in each industry that provides appreciable information. Each of these studies consolidated and/or more accurately refined the populations used in the earlier studies. This needs to be clarified 6 because it is central to the understanding of how consistent the relationship is between 1,3 butadiene and cancer. For example, EPA should explain that the 8 Delzell study included all of the eligible population from the Matanoski (1990) 9 study and the Meinhardt study (1982). The leukemia finding in the Matanoski et al. 10 (1990) study should not be presented as if it were a separate finding in a completely different population from the Delzell study. In addition, the Delzell 12 study supersedes this and other previous studies and rectifies many of the 13 limitations and errors of the earlier studies. 15

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- c) Page 11-5, line 12-34: The Matanoski study is presented as if it were a study on a completely different population from the Delzell study. In discussing the excess leukemia risk in the nested case-control study in the Matonoski study, it is important to indicate in line 14 that there was no excess of leukemia observed in the cohort study (standardized mortality ratio was 1.0 representing 22 observed, 22.9 expected). This is an important major point to be made up-front in this section because it helps explain the scientific debate later referred to in lines 26-28. The evidence linking butadiene exposure and cancer is still strongest for leukemia based on one large, high quality cohort study of SBR workers (Delzell studies) which supercedes the Matanoski study.
- Pages 11-6 to 11-7: It is never clearly stated that the leukemia excess seen in the d) Delzell study has not been replicated in a completely different study population. The studies of butadiene monomer workers and of other butadiene exposed

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ii)

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1 workers report null results for leukemia. Page 11-7, line 17: The lymphosarcomas and leukemias are discussed as if they 2 e) 3 could be considered as the same type of cancer. It is implied that two different 4 populations have an excess in lymphohematopoietic cancers thereby demonstrating 5 consistency across studies. The strength of association evaluation should focus on 6 these different cancers separately. The EPA review states on page 11-7, lines 20-26 that the monomer workers 7 8 exposed to shorter periods of time probably had higher exposures than workers 9 exposed for longer periods of time. This statement needs to be removed as there is 10 no evidence for this. The available evidence is insufficient for a causal relationship 11 between butadiene and lymphosarcoma. Lymphosarcoma was elevated for short 12 term exposed workers but not among long term exposed workers indicating a lack 13 of dose-response. Of equal importance, lymphosarcoma/non-Hodgkins lymphoma 14 (NHL) was not elevated in the SBR cohort which had excess leukemia. These 15 points must be discussed in the EPA document. The possibility that it is the SBR 16 process and not butadiene alone that may explain excess leukemia should be 17 discussed regardless of final decision on cancer classification. Delzell's finding that 18 the leukemia excess concentrated among workers who began employment in the 19 1950's and not those that worked exclusively in 1940's led to hypothesis by Irons 20 and Pyatt (1998) that DMDTC might be a contributing factor in the leukemia 21 excess. This hypothesis needs to be discussed as it is part of the scientific 22 literature. 23 f) Page 11-8, Table 11-2: Some of the entries in this table seem unbalanced. 24 i) Matanoski is cited as showing "7 to 9 times higher relative odds for 25 leukemia" without mentioning that others analyzed the same data using a

slightly different cutpoint and found an odds ratio of <1.0.

Most of the dose-response tabulations by Delzell et al. 1996 do not show

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1		mortality ratios in the highest dose groups as high as the ones cited.
2		iii) It states that a dose-response relation for monomer plant workers "Cannot
3		be demonstrated due to lack of quantitative exposure data," when in fact
4		several dose-response analyses were performed (albeit with an imperfect
5		dose metric) and showed not even a hint of an association.
6	g)	Page 11-9, Line 24: The Committee classified the SBR process as a known human
7		carcinogen.
8	h)	Section 11.3.3, Page 11-9: This section needs to more accurately reflect the
9		scientific literature that demonstrates a clear difference in metabolic activation
10		between humans and rodents and should discuss how PBPK modeling can refine
11		the risk assessment process. If EPA cannot incorporate PBPK modeling into risk
12		assessment within the mandated time-contraints, EPA should discuss future
13		possible directions and consider alternative ways (reduced safety factor) to account
14		for the species differences.
15	i)	Page 11-10, line 36 to Page 11-11, lines 1-2: The EPA states that the tumor type
16		in rodents most analogous to the lymphohematopoietic cancers is the lymphocytic
17		lymphomas. To properly discuss the strengths and weaknesses of this statement,
18		EPA should discuss the data generated by Irons et al, 1996 which shows that T-
19		cell lymphoma in mice is due to a specific population of stem cells in the mouse
20		bone marrow that is not present for humans or rat bone marrow cells. EPA should
21		also point out that the link between 1,3-butadiene exposure and lymphoma is weak
22		as there was no consistent dose-response relationship. Additionally, the Agency
23		should show consistency in combining different tumor types across documents. For
24		example, in the revised cancer risk assessment guidelines a case study for an
25		aromatic hydrocarbon (presumably benzene) is presented where mice are shown to
26		develop lymphomas following exposure whereas humans develop acute
27		myelogenous leukemia. In this case, the Agency did make the distinction that the

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1 response in the animal models and the response in the humans was different. The 2 Agency should acknowledge that there is considerable disagreement as to whether 3 these two tumor types, lymphomas and leukemias, derive from the same origin. 4 The potential impact of task specific peak exposures must be addressed. The 5 primary metric for SBR workers is based on time weighted average exposure. But 6 SBR workers frequently get the majority of their exposures during a small fraction 7 of the work day during the conduct of specific tasks. The leukemogenic effect was 8 associated with those jobs that involved high peak exposures in latex sampling 9 (laboratory workers and in vessel cleaning for maintenance laborers). The general 10 population is typically exposed to lower ambient levels. An additional uncertainty 11 that should be discussed is the uncertainty of extrapolating from an occupational 12 setting where peak exposures occurred to the generally low ambient levels of 13 exposure. 14 j) Page 11, Lines 11-13: The statement that the evidence regarding human 15 carcinogenicity is based on ten studies is deceptive, because the reports are not 16 independent, but most reports are updates of previous ones. In addition, there are 17 basically only four independent cohorts, not five, because the Delzell study 18 included about 95% of the workers in the Matanoski studies. 19 Section 11.5, Pages 11-13-11-14: This section needs to be re-evaluated in light of k) 20 the two more recent negative dominant lethal studies. A weight of evidence 21 approach should be taken looking at all 3 studies rather than to emphasize the one 22 positive study. 23 1) Section 11.5 Pages 11-14, lines 1-3: This sentence should be rewritten as fact not 24 as speculation. 25 Pages 11-14, lines 10-16: These two sentences should be rewritten or removed. m) 26 Text discusses concept of "meaningful increases in risk." What is the definition of

this phrase? Is this in the USEPA guidelines? This phrase appears to be an

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1		editorial. For this reviewer to understand that "effects are not expected in
2		humans exposed to low environmental exposures " the assessment must provide
3		a detailed rationale that follows from Chapters 5 and 9. This summary statement
4		was not supported in the document.
5	n)	Section 11.6, Pages 11-15: This summary section needs to be integrated with the
6		biology discussed in Chapter 5 and quantitative assessments in Chapter 9. No new
7		"surprise," non-justified statements should appear in this section.
8	o)	Section 11.6: The importance of the positive heritable translocation studies in mice
9		should be more heavily emphasized.
10	p)	Section 11.6, Page 11-15, line 10-17: The additional safety factor of 3 to
11		extrapolate from a LOAEL to NOAEL is inappropriate and should be removed.
12		Inadequate justification is given for the application of the additional safety factor
13		for the benchmark dose. The EHC could not understand the rationale for its
14		inclusion. This section must discuss the rationale for using the "hybrid model" for
15		continuous data analysis and exactly what the hybrid model is so that the risk
16		assessment can be completely transparent. On the surface it appears that the
17		approach yielding the lowest LEC was selected. Some of the EHC
18		Members/Consultants recommend that the Agency use the EC10 (central estimate)
19		as the point of departure rather than the LEC10. Other EHC Members/Consultants
20		did not agree with the recommendation.
21	q)	Section 11.8 on Page 11.16
22		The Agency should expand on future research needs to fill gaps in knowledge. The
23		list is inadequate.
24		
25	3.2	Classification of 1,3-Butadiene as a Known, Human Carcinogen
26	Does	the science support the classification of "known" human carcinogen?
27	There	was a majority opinion amongst the Committee that the science supports the SBR

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The case for an association between lymphosarcoma and butadiene exposure is weakened by the fact that in the main study (Divine, et al., 1996) that purportedly showed the association, there

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was no indication of an exposure-response relationship nor was there evidence that those with longer-term exposure had a higher risk of lymphosarcoma.

On page 11-3, lines 11-13, the report claims that "'sufficient evidence' of human carcinogenicity is based on more than 10 epidemiological studies examining five different groups of workers" and summarizes them in Table 11-1. But there are effectively only 4 groups of workers, not 5, since the Matanoski and Delzell cohorts have a high degree of overlap (i.e., about 95% of the Matanoski cohort is included in the Delzell cohort). In addition, the "more than 10" studies are just earlier reports of the same cohorts and do not add anything to an inference of causality beyond that seen in the most recent follow-ups. Of the four independent studies, one small one Cowles, et al., 1994 is completely negative for lymphosarcoma and leukemia; another small one was positive for lymphosarcoma, but based on only 4 cases, and showed no excess of leukemia (Ward et al., 1996); a relatively large study was mostly negative, but a bit suggestive, of a lymphosarcoma excess, but provided no support for a leukemia excess (Divine, et al., 1996); and one large study was positive for leukemia but not for lymphosarcoma (Delzell, et al., 1996).

In summary, the weight of epidemiological evidence does not support an association between butadiene exposure and lymphosarcoma/reticulosarcoma.

While the Delzell study (Delzell, et al., 1996, Macaluso, et al., 1996) is a large and methodologically sound study, one would like to see at least a second independent confirmatory study before affirming there is "sufficient evidence of human carcinogenicity" regarding butadiene and leukemia. Instead, one sees a fairly large and reasonably sound study that shows no leukemia excess (Divine, et al., 1996) plus two smaller ones with no evidence of leukemia risk, and these weaken the case.

A majority of the EHC members felt that the body of mechanistic data on butadiene does not support the classification of known human carcinogen, while others did not. The Agency notes (page 9-52) that there are large unexplained differences in the response of rats and mice to butadiene and states that the specific mechanisms of 1,3-butadiene induced carcinogenesis are

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unknown. However, there is strong evidence from both metabolism and genetic toxicology studies that diepoxybutane is a critical metabolite in the carcinogenic process. The extensive capability of mice to form the diepoxide metabolite and the extensively higher levels of the diepoxide metabolite in blood and tissues of mice compared with rats, most likely forms the basis for this dramatic species difference in carcinogenic response. This is an important point because it suggests that the diepoxide metabolite may be the best dosimeter for assessing risks for humans exposed to butadiene.

Toxicokinetic modeling cannot prove or disprove the relationship between chemical exposure and toxic or carcinogenic effect. However, these models can be developed to test quantitative hypotheses regarding proposed mechanisms of toxicity or carcinogenicity of chemicals. The Agency notes that "pharmacokinetic modeling of 1,3-butadiene has not elucidated the reasons for interspecies differences in carcinogenic response between rats and mice" and "mice and rats also exhibit substantial quantitative differences in their metabolism of 1,3-butadiene to potentially reactive metabolites. Unfortunately, existing pharmacokinetic models have been unable to explain the species differences in carcinogenic response." These statements are not true if one considers the current PBPK models that describe the disposition of butadiene, epoxybutene and diepoxybutane. These PBPK models clearly simulate the dramatic species differences in tissue and blood concentrations of diepoxybutane between rats and mice observed in vivo. The models suggest that species differences in response to butadiene are most likely related to differences in rates of formation and removal of diepoxybutane. Experimental data and PBPK model simulations indicate that mice produce far greater concentrations of this reactive metabolite compared with rats.

These PBPK models can be, and have been, extended to humans. In vitro data on rates of butadiene, epoxybutene, and diepoxybutane metabolism obtained in human tissue samples can also be used to predict blood and tissue concentrations of both epoxide metabolites in humans. When these simulations are conducted using average values for human metabolic rates results indicate that human diepoxide concentrations would be orders of magnitude less than those of

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mice and lower but much more similar to concentrations predicted for rats. Nonetheless, 1,3-butadiene produces cancer in rats, albeit at higher dose levels than for mice, so similarity of human and rat metabolism would still contribute to the weight of evidence. Some Committee members point out limitations in the models and the underlying assumptions regarding choice of human parameters and activity of metabolites. These members also point to the considerable other mechanistic data (which are described in comments on Chapter 4).

3.3 Approaches Taken to Characterize Plausible Cancer Risks

Are the approaches taken to characterize plausible cancer risks reasonable given the science? Butadiene is one of the first chemicals to be brought forward under the EPA's new risk assessment guidelines. It needs to be a model of how to proceed. The Agency must clearly separate known from unknown and important from unimportant. The database for butadiene is very challenging and robust, but all of the important issues are not settled. The EPA must state the facts in proper perspective and should not be afraid to say that more information is needed.

Risk estimates use external butadiene exposures as the dose estimate given the sophisticated PBPK models currently developed and the availability of in vitro metabolic parameters in rodents and humans an approach that used an estimate of internal dose. The Agency notes (Page 11-12, Lines 1-3) that a review of the available pharmacokinetic data and models reveal that the state of this science is currently inadequate for either explaining interspecies differences or improving on default dosimetry assumptions. As noted previously in this report, this statement should be revised to reflect the available scientific data and pharmacokinetic models. The fact that human variability in metabolic response is noted should not

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be viewed as a limitation for these models, but instead as an opportunity to reduce uncertainty and characterize variability. If metabolic activation of butadiene is necessary to produce genetic damage, and ultimately carcinogenicity, which is not an unreasonable hypothesis, then individuals with the highest metabolic capacity for activation and the lowest metabolic capacity for deactivation would be at most risk for exposure to butadiene. The PBPK model could be used to determine the extent to which metabolic differences among humans is the dominant process controlling the magnitude of the effective dose at the target site.

The Agency notes (page 9-16, lines 1-7) that risk assessments based on rat carcinogenicity data "are not considered the most appropriate estimates of human risks EPA believes that the mouse is likely to represent a better rodent model for human cancer risk assessment from 1,3-butadiene." Available mechanistic data on the formation of epoxybutene and diepoxybutane obtained in rat, mouse and human tissue samples suggest that the rat is a more appropriate model for assessing risks for humans than is the mouse. Choosing a rat versus a mouse as the most appropriate animal for assessing risks in humans may, in fact, be an oversimplification. As noted above, the development of PBPK models that are capable of predicting concentrations of the reactive metabolites in the target tissues and availability of distributions of human metabolic rate constants would allow one to not rely on either mouse or rat per se. Instead one can use all of the available mechanistic data within the context of a PBPK model to predict the butadiene doses in humans necessary to yield epoxide concentrations in tissues similar to those predicted for rats or mice exposed to carcinogenic concentrations of butadiene.

The Agency notes (page 9-17, lines 6-11) that no attempt was made to adjust for internal doses of reactive 1,3-butadiene metabolites because the PBPK data were inadequate to develop a reliable PBPK model. As noted previously PBPK models not reviewed by the Agency in this draft document have been developed that are capable of predicting the butadiene epoxybutene and diepoxybutane blood and tissue concentrations in rats, mice and humans following exposure to butadiene. In this case, a human PBPK model could be readily used to obtain either point estimates using average human values or a range of estimates by conducting Monte Carlo simulations to

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sample from a distribution of available parameter values for humans for both metabolic rate parameters and physiological parameters. A plausible assumption is that for a given diepoxybutane tissue concentration the human response would be equivalent to the rodent response. Then, concentrations of butadiene necessary to elicit tissue doses of diepoxybutane in humans equivalent to diepoxybutane concentrations in mice or rats at doses which yielded tumors could be calculated. PBPK models of these and the diepoxybutane and other active metabolites can then be applied to human risks predictions. The advantage of using a PBPK model is that mechanisms underlying absorption, distribution, metabolism and elimination of butadiene, epoxybutene, epoxybutanediol and diepoxybutane which are either similar or different across species and across doses would be accounted for. Given the availability of these PBPK models, this approach would be far superior to using a simple arithmetic adjustment for continuous daily exposure versus exposure under bioassay conditions.

On page 9-25, lines 1-7 the EPA notes that "mice and rats also exhibit substantial quantitative differences in their metabolism of 1,3-butadiene to potentially reactive metabolites. Unfortunately, existing pharmacokinetic models have been unable to explain the species differences in carcinogenic response." A majority of the Committee assert that there are several published PBPK models that have been able to successfully explain species differences in the carcinogenic response exposure to 1,3-butadiene based on differences in the metabolism of 1,3-butadiene to reactive metabolites while other Committee members were concerned about the degree to which some models were able to describe the available data, the extent to which parameters had to be changed from measured values to obtain reasonable fits and the accuracy of the low dose extrapolation. In particular, the highly mutagenic diepoxybutane is formed to a much greater extent in mouse tissues compared with rats.

Again on page 9-25, lines 15-19, the EPA notes that "ideally a PBPK model for the internal dose of the reactive metabolites would decrease some of the quantitative uncertainty in interspecies extrapolation. However, current PBPK models are inadequate for this purpose." The available

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pharmacokinetic models and emerging data on human pharmacokinetics should be further explored for use in risk assessment. As noted in the previous comments, some of the Committee members assert that PBPK models that are capable of interspecies and low dose extrapolation for this purpose do exist. Other Committee members assert that although the PBPK models require validation and development they may prove useful in the near term and that models should be further developed to address parameter and structural uncertainty as well as interindividual variability in human metabolism, but the tools for such activity are at hand. On page 9-51 the Agency notes that NOAEL LOAEL EC10 or LEC10 should be converted to appropriate human equivalent exposures before using these exposure levels as points of departure and that theoretically this is best accomplished using a PBPK model. The Agency also notes that the current PBPK models are inadequate for use in risk assessment. However, as noted previously, some of the Members felt that there are several available PBPK models that are capable of predicting diepoxide concentrations in target tissues for rats, mice and humans and that any one of these models could be used to obtain a more appropriate human equivalent exposure. Other Committee members felt that, at a minimum, the different toxicokinetic hypotheses and the hypothesized role of the various metabolites should be discussed, at least qualitatively, with an indication of the degree to which the assessment would be impacted if some of the hypotheses were later proven true. In the 1,3-butadiene health risk assessment, the Agency should address the variety of viewpoints and hypotheses of research in the field of PBPK modelling.

The EPA report pays relatively little attention to the issue of how much peak exposures to butadiene may have influenced leukemia risk in the Delzell study. It indicates (P. 7-22, L. 30-33) that there was an association of peak exposures (defined as >100 ppm) with leukemia but dismisses it as an "irregular" association. The Acquavella commentary on the EPA draft that was sent to the EHC (Acquavella, 1998) presents a table showing how much peak exposures affect the association of butadiene ppm-years with leukemia risk (Page 10). The formal linear regression

estimates with and without adjustment for peak exposures are not presented, but it is apparent that the regression estimate would be appreciably less when peak exposure was adjusted for. Specifically, for the three highest cumulative exposure groups, the excess relative risks with and without control for peak exposures were 0.0 and 1.0 respectively for 20-99 ppm-years, 0.3 and 1.4 respectively for 100-199 ppm-years, and 1.5 and 3.6 respectively for 200+ ppm-years. In each dose group, adjustment for peak exposures reduced the leukemia risk substantially. Since butadiene exposures to the public will almost never approach the peak exposure range, a more appropriate model for risk would factor out the peak-exposure component.

On Page 9-2, Lines 24-25 regarding the Delzell analysis of butadiene exposure vs. leukemia, it is noted that "excluding exposures within 20 years of death weakened and almost eliminated the relationship...." This indicates that in modeling lifetime risk, a model that assumes a limited effect time (i.e., that leukemia risk during a given year of age is affected largely by the butadiene exposures received during the previous, say, 20 years, and only slightly or not at all by more distant ones) should be considered. This "windows of exposure" model has precedents, e.g., lung cancer risk from radon has been modeled in this way in a National Academy of Sciences report (NAS/NRC, 1988) because lung cancer risk was little affected by radon exposures in the distant past; leukemia risk from radiation is highly elevated at 5-10 years after irradiation but there is little elevation by 20-30 years after irradiation (NAS/NRC, 1990). If this model were considered for projecting lifetime risk, it would show appreciably less risk from chronic exposures than does the present one, which assumes that excess relative risk at, say, age 70 is an additive function of all the exposure accumulated in the previous 69 years.

3.4 Conclusions and Quantitative Estimations for Reproductive/Developmental Effects

Are the conclusions and quantitative estimations for reproductive/developmental effects adequately supported?

The Committee supports the Agency's use of benchmark dose procedures and the

modeling of reproductive toxicity endpoints. The EHC also supports the continuing attempts to						
develop new strategies, such as those presented in the report to address ovarian, uterine and						
testicular atrophy, to quantitatively address reproductive endpoints in risk assessment documents.						
This is one of	the first such assessments and some general suggestions are made regarding conduct					
of the analyse	es and the presentation of results for the analyses:					
a)	A clear rationale for the selection of one particular point of departure over another is					
	needed for each case for which the default is applied and for cases deviating from					
	the default.					
b)	Where it is not obvious the biological significance of particular endpoints for the					
	assessment should be conveyed.					
c)	When applying novel approaches, such as the time-to-response modeling of					
	atrophy, the advantages and limitations in utilizing the approach in the context of					
	reference concentration estimation should be clearly understood and presented.					
d)	The incorporation of information on toxicokinetics (e.g., ovotoxicity of diepoxide)					

1		should be used where possible on at least a qualitative if not quantitative basis to
2		inform the choice of endpoints and modeling.
3	e)	The exclusion of points at high doses contributing to a poor fit is a reasonable
4		approach, and typically can be explained, for example, in terms of toxicities other
5		than the one being modeled, or toxicokinetics. When the approach is applied where
6		possible the specific rationale for the case at hand should be presented. The use of
7		the procedure and the rationale provided for should be consistent across endpoints
8		and across assessments for the same scenarios.
9	f)	There may be a difference of opinion among Committee members about the
10		appropriateness of using a risk reduction factor, when reference concentrations are
11		based on benchmarks associated with risk of effect. The rationale for use of this factor
12		needs to be clearly laid out.
13	g)	Analyses of this type applied to these endpoints may be unfamiliar to others
14		working in the general area. Confusion over the presentation of some results
15		(e.g., logarithm versus linear scales) was obvious in some of the comments
16		received. Also, calculations should be easily followed, and of course need to be
17		carefully proofed by someone other than the one making them. Tables and figures
18		should be free standing, with statistics referenced.
19	Some	of the Committee members are of the opinion that the quantitative estimations for
20	reproductive/o	developmental effects could benefit greatly from the application of PBPK modeling to

estimate the effective dose at the target site. The Agency notes (Page 9-46, lines 6-11) that

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e)

"ovarian atrophy has been shown to be related to the amount of the diepoxide metabolite in the						
tissue. Modeling of the ovarian atrophy and uterine atrophy data was considered based on internal						
dose of the diepoxide metabolite, however an adequate model was not available to estimate levels						
of the diepoxybutane." There are several PBPK models that are currently published that allow						
calculation of the dose of the epoxide metabolite in target tissue. Any one of these models could						
be used to determine the internal dose of the diepoxide metabolite and provide a more refined						
estimate of risks for this reproductive endpoint.						
4. SUMMARY OF RECOMMENDATIONS						
a) The EHC recommends that the Agency's updated, draft health risk assessment of						
1,3-butadiene reflect the new information that has been published in the peer-reviewed						
literature since the Agency's cut-off date of January 31, 1997. The inclusion of this						
new information would greatly improve the technical quality and comprehensiveness of						
the report.						
b) Since critical research like the Delzell et al. (1995) exposure reestimation and the						
pharmacokinetic modeling has not been completed yet, the Committee recommends						
that the Agency's risk assessment of 1,3-butadiene be labeled as an interim risk						
assessment.						
c) The majority of the Committee recommends that the Agency classify 1,3-butadiene as a						
probable human carcinogen because there was not consistency between the exposure and						
leukemia or lymphosarcoma data when the SBR and monomer studies were considered in						
total. Also, the role of confounders and cofactors was unclear.						
d) The Committee found the approaches taken to characterize plausible cancer risks to be						
reasonable but points out specific improvements that can be made.						

The Committee supports the Agency's use of benchmark dose procedures and the

- development of mathematical models for reproductive endpoints. To improve the analysis
- and the clarity of the results specific recommendations focused on:

1		(i) providing a clear rationale for selecting one particular point of departure
2		over another,
3		(ii) conveying the biological significance of particular endpoints of the
4		assessment,
5		(iii) presenting the advantages and limitations of novel approaches such as the
6		time-to-response modeling of atrophy,
7		(iv) incorporating information on toxicokinetics on at least a qualitative, if not
8		quantitative basis to inform the choice of endpoints,
9		(v) excluding points at high doses that contribute to a poor fit,
10		(vi) explaining the rationale for using a risk reduction factor, and
11		(vii) proofing calculations and correcting them where needed.
12	f)	In Chapter 1, Introduction, the Agency should state whether the different cancer
13		classification systems and quantitative assessments are equally valid and
14		scientifically defensible and explain the rationale. In addition, the Agency should
15		comment on its Cancer Risk Assessment Guidelines.
16	g)	In Chapter 2, Overview of Exposure to 1,3-Butadiene, the Agency should clearly
17		indicate that the chapter is not intended to be a comprehensive review of exposure.
18		The chapter should include an explanation on how concentrations have been
19		measured over the years and how this might impact a comparison of exposure
20		levels measured in earlier years with those from more recent years. Specific
21		recommendations for improvements for chapter 2 are included in Section 3.1.2.

1	h)	There are sev	eral recommendations regarding Chapter 3, Metabolism and
2		Pharmacokine	etics. The main recommendations include the following with additional
3		comments inc	eluded in Section 3.1.3.
4		i)	The title of Chapter 3, Metabolism and Pharmacokinetics, should be
5			changed to Metabolism and Toxicokinetics because 1,3-butadiene is not
6			used as a therapeutic agent.
7		ii)	The word, toxicokinetics, should replace the word, pharmacokinetics,
8			throughout the document.
9		iii)	Many new studies are not incorporated into the chapter. The Health
10			Canada assessment and the comments that were submitted by Dr.
11			Himmelstein provide many of those new references (Himmelstein, 1998).
12		iv)	The Agency should revisit its statement regarding species differences in
13			butadiene metabolism, taking into account the most recent information on
14			species differences in the production of diepoxybutane and other reactive
15			metabolites.
16		v)	The chapter should point out the close parallels between the observations in
17			in vitro studies on metabolism and tissue concentrations of epoxybutene and
18			diepoxybutane in mice and rats and in vivo studies on the metabolism of
19			butadiene.
20	i)	The C	committee's recommendations for Chapter 4, Mutagenicity, are numerous.

1	Some	of the recommendations are given below and additional recommendations
2	are inc	cluded in Section 3.1.4:
3	i)	The statement on Page 11-1 regarding the lack of sufficient data to
4		determine if children or other subpopulations are afffected differently by
5		exposure to 1,3-butadiene should be revisited in light of the studies by
6		Nelson et al., 1995, Wiencke et al. and Kelsey et al, 1995. Nelson et al.,
7		1995 found the glutathione transerase theta (GSTT1) to be highly
8		polymorphic wide variation in its ethnic distribution (Nelson et al., 1995).
9	ii)	Most of the extensive work on mutagenicity prior to 1994 should be
10		included.
11	iii)	The chapter should include text tables that summarize key animal and
12		human findings derived from the entire body of information on butadiene.
13	iv)	The tables should include references to support key findings.
14	v)	There should be separate tables for in vitro, animal and human findings.
15	vi)	The similarities and species differences in response should be noted.
16	vii)	The chapter should include more emphasis on the positive heritable
17		translocation studies in mice because of their potential relevance for human
18		heritable risks, and to several additional studies conducted in humans.
19	viii)	The conclusion section should be expanded to include what is known about
20		the mutagenicity of 1,3-butadiene and its metabolites.
21	ix)	The missing dose units, missing units for mutant frequencies and other

1			similar omissions should be added.
2		x)	The explanation as to possible reasons for the discordance between the
3			positive effects obtained by autoradiography and the negative results found
4			by cloning for human 1,3-butadiene induced hprt mutations in vivo should
5			be rewritten.
6		xi)	The conclusion section should be expanded by adding statements
7			summarizing what is known about mutagenicity of 1,3-butadiene and its
8			metabolites.
9	j)	Some	of the recommendations for Chapter 5, Reproductive and Developmental
10		Effects	s, are included in recommendations cited in (e). Additional recommendations
11		are pro	ovided in detail in Section 3.1.5 and are summarized below.
12		i)	All of the pertinent studies should be introduced in a summary table at the
13			beginning of each section.
14		ii)	Both positive and negative studies should be included so the reader can
15			develop a comprehensive understanding of the data base supporting the
16			assessment.
17		iii)	More current research should be included, especially the new dominant
18			lethal studies.
19		iv)	Chapter 5 should be integrated with the other chapters, especially the

- DO NOT CITE OR QUOTE -1 chapters on pharmacokinetics and metabolism, animal toxicity, and 2 quantitative risk assessment. 3 The Agency should identify where it is uncertain with regard to conclusions v) about the reproductive and developmental endpoints, identify the action it 4 5 will take to respond to the uncertainty, and should include all of the assumptions regarding uncertainty in the respective chapter. 6 7 The Committee's recommendations on Chapter 6, Toxicity in Animals, are listed in k) 8 Section 3.1.6 and include the following: 9 i) The rationale for the selection of the toxic non-cancer endpoint that is 10 utilized in the derivation of the RfC is very important and should be more explicitly 11 explained. 12 ii) It is unclear whether all of the repeat dose studies have been reviewed by 13 the Agency. The EPA should incorporate the repeat dose in vivo 14 mammalian studies of 1,3-butadiene in Chapter 6 unless these are covered 15 elsewhere in the document and the Agency explicitedly states so. 16 The Agency should present the NTP study details in the first section iii) 17 (chronic) of Chapter 6 and then refer to them in the carcinogenicity section. 18 19 iv) The EPA should use the same categories of tumors for presenting data

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from the continuous treatment, 9 and 15-month interim sacrifices and stop-

1			exposure study in order to permit comparisons.
2		v)	A table summarizing the positive oncogenic findings across all studies
3			specifying the dose tested, and the type of tumor that was significantly
4			elevated, should be added since there were numerous organs and tumors
5			involved.
6		vi)	The carcinogenic evidence on the mammalian metabolites of 1,3- butadiene
7			should be separated from the data on related chemicals and placed in a
8			subsection by itself.
9		vii)	The rat carcinogenicity data should be presented in similar detail to the
10			NTP data so that the reader does not have to find the old risk assessment
11			to see the data.
12		viii)	The observation that concentration, not time is a critical determinant of
13			potency is not supported by a comparison of the tumor data presented in
14			Table 6-4 and Table 6-8 from the NTP (1993) study for continuous lifetime
15			treatment and stop-exposure study.
16	1)	The C	Committee recommendations for Chapter 7 include:
17		i)	A statement that, for the Delzell study (1996), there was no excess among
18			those hired before 1950 (Observed/Expected = 17/16.4 = 1.04) when one
19			would expect the highest exposures, but there was an excess among those
20			hired during 1950-59 (Observed/Expected = 20/10=2.0, Confidence

1			Interval = $1.2-3.1$)
2		ii)	A statement regarding the inappropriateness of "lumping" the
3			lymphohematopoietic tumors should be added.
4		iii)	A statement regarding the possible role of confounding should be included
5			in the document.
6	m)	The recomme	endations for Chapter 8 are provided in Section 3.1.8 and make the following
7		revisions:	
8		i)	The most recent relevant literature on butadiene toxicokinetic modeling
9			should be included.
10		ii)	The Agency should revisit its conclusion in Chapter 8 once it includes the
11			recent PBPK models as explained in Section 3.1.8.
12	n)	The recomme	endations for Chapter 9 are provided in Section 3.1.9 and include the
13		following:	
14		i)	The 1997 data of Matanowski, et al. should be considered,
15		ii)	The Agency should explain or provide further explanation on the following:
16			a) why equivalent model fits for the Delzell data are obtained in order
17			to convey a greater appreciation of the nature of the dose response
18			data,

1		b)	the role of benzene as a confounder,
2		c)	the potential importance of the uncertainties of dose reconstruction
3			and the potential magnitude of the impact on the risk predictions,
4		d)	where possible, the rationale for the exclusion of points at high
5			doses contributing to a poor fit, and
6		e)	information on toxicokinetics (e.g., ovotoxicity of diepoxide).
7	o)	Chapter 10, V	Veight of Evidence, should be rewritten to reflect the range of
8		opinion regard	ding the human findings. In addition, the finding that 1,3-butadiene,
9		when mutage	nic or clastogenic, is so through its metabolism should be provided in
10		both the evalu	nation and in the statement regarding the mechanistic studies that are
11		most relevant	to humans.
12	p)	Specific recor	mmendations for Chapter 11, Risk Characterization, are provided in
13		Section 3.1.1	1 and include the following:
14		i)	The statement that the conclusion of "sufficient evidence" of human
15			carcinogenicity is based on more than 10 epidemiologic studies
16			examining five different groups of workers should be rewritten
17			since it is misleading.
18		ii)	In discussing the excess leukemia risk in the nested case-control

1		study in the Matanoski study, it is important to indicate that there
2		was no excess of leukemia observed in the cohort study.
3	iii)	The chapter should clearly state that the leukemia excess observed
4		in the Delzell study has not been replicated in a completely different
5		study population. In addition, a weight of evidence approach should
6		incorporate all three studies rather than to emphasize the one
7		positive study.
8	iv)	The strength of the association evaluation for the lymphosarcomas
9		and the leukemias should focus on these different cancers
10		separately.
11	v)	The EPA should incorporate the PBPK modeling into Chapter 11 if it
12		has time to do so or should at a minimum, discuss future possible
13		directions and consider alternative ways (e.g. reduced safety factor)
14		to account for the species differences.
15	vi)	Some members felt that the additional safety factor of 3 to move
16		from an effect dose to a no effect dose should be removed because
17		it is inappropriate.
18	vii)	The document should point out important data gaps in our
19		knowledge and research needs.

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1 2	<u>A</u>	PPE	NDIX A - ACRONYMS AND ABBREVIATIONS
3 4	ACGIH-TLV	-	American Conference of Governmental Industrial Hygienists - Threshold Limit Value
5	Bdiol	-	3-butene-1,2-diol
6	CI	-	confidence interval
7	DEB	-	1,2,3,4-diepoxybutane
8	DMDTC	-	Dimethyldithiocarbamate
9	EB	-	1,2 epoxy-3-butene
10	EBD	-	3,4-epoxy-1,2-butanediol
11	Ec_x	-	effective concentration at x% risk, ranging from 0.1% to 10%
12	EHC	-	Environmental Health Committee
13	ED	-	effective dose
14	EBdiol	-	1,2 dihydroxy-3-4 epoxybutane
15	GC-MS	-	gas chromatography - mass spectrometry
16	GSTT1	-	glutathione S-transferase theta
17	GSTM1	-	glutathione S-transferase μ
18	HEI	-	Health Effects Institute
19	ICD	-	International Classification of Diseases
20	JEM	-	job-exposure matrices
21	K_{m}	-	substrate concentration at one-half maximum velocity

DRAFT #4 - DRAFT DOCUMENT-- DO NOT CITE OR QUOTE -1 LEC_p 95% lower confidence intervals associated with a risk (p), ranging from 2 1% to 10% 3 LOAEL - lowest-observed-adverse effect 4 MACT maximum achievable control technology milligram 5 mg maximum likelihood estimate 6 MLE 7 NCI National Cancer Institute 8 NHL non-Hodgkins lymphoma NOAEL no-observed-adverse effect 10 NTP National Toxicology Program 11 O/E observed/expected Office of Mobile Sources 12 OMS 13 ORD Office of Research and Development 14 **PBPK** physiologically-based pharmacokinetic 15 parts per million ppm 16 parts per million per hours ppmh 17 the incremental unit cancer risk for humans q_1 18 RfC Inhalation Reference Concentration 19 RR relative risk 20 SBR styrene butadiene rubber

sister chromatid exchange

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SCE

DRAFT #4 - DRAFT DOCUMENT-- DO NOT CITE OR QUOTE -1 SOT Society of Toxicology 2 THB trihydroxybutane TRI Toxics Release Inventory 3 4 UAB University of Alabama at Birmingham maximum velocity for an enzyme-mediated reaction 5 V_{mzx} 6 WWII World War II

- DRAFT DOCUMENT-- DO NOT CITE OR QUOTE -

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